

INVESTIGATIVE REPORT

Microvessel Density and Expression of Vascular Endothelial Growth Factor and its Receptors in Different Subtypes of Primary Cutaneous B-cell Lymphoma

Marion WOBSE¹, Claudia SIEDEL¹, Hermann KNEITZ¹, Eva-Bettina BRÖCKER¹, Matthias GOEBELER¹, Roland HOUBEN^{1#} and Eva GEISSINGER^{2#}

¹Department of Dermatology, Venereology and Allergology, and ²Institute of Pathology, University of Wuerzburg, Wuerzburg, Germany

[#]These authors contributed equally.

A proangiogenic micromilieu is associated with a worse prognosis in systemic lymphoma. Hence, targeting the tumour microenvironment and its vasculature has evolved as a promising novel treatment strategy. The role of tumour neoangiogenesis in cutaneous B-cell lymphoma, however, has not yet been elucidated. Therefore, we examined the expression of vascular endothelial growth factor (VEGF) and its receptors VEGFR-1 and VEGFR-2, as well as microvessel density by immunohistochemistry in paraffin-embedded specimens of different subtypes of primary cutaneous B-cell lymphomas, systemic diffuse large B-cell lymphoma, and cutaneous B-cell pseudolymphoma. Primary cutaneous large B-cell lymphoma (PCLBCL) were characterized by significantly higher intratumoral expression levels of VEGF and its receptors in comparison with the indolent lymphoma subtypes. Moreover, PCLBCL exhibited significantly higher intratumoral microvessel counts. Our study provides evidence that the most aggressive subtype of cutaneous B-cell lymphoma, PCLBCL, is characterized by a proangiogenic micromilieu. Key words: angiogenesis; microvessel density; VEGF; cutaneous B-cell lymphoma; tumour microenvironment.

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Marion Wobser, Department of Dermatology, Venereology and Allergology, University of Wuerzburg, Josef-Schneider-Str. 2, DE-97080 Wuerzburg, Germany. E-mail: Wobser_M@klinik.uni-wuerzburg.de

According to the World Health Organization/European Organisation for Research and Treatment of Cancer (WHO/EORTC) classification, primary cutaneous B-cell lymphomas (PCBCL) are subdivided into 3 major groups; namely, primary cutaneous follicle centre lymphoma (PCFCL), primary cutaneous marginal zone lymphoma (PCMZL), and primary cutaneous large B-cell lymphoma (PCLBCL) (1). While PCFCL and PCMZL show an indolent clinical course with an excellent prognosis, PCLBCL represents a more aggressive lymphoma. In contrast to diffuse PCFCL, in which

small to large cleaved centrocytes dominate the tumour infiltrate, PCLBCL is histologically characterized by large sheets of centroblasts and immunoblasts. Age and tumour extent represent negative prognostic factors in PCLBCL. Moreover, impaired prognosis is observed when skin lesions are present at the lower extremities, and this has given the name to this lymphoma subgroup PCLBCL, leg-type (PCLBCL-LT)), a hitherto incompletely understood phenomenon. Similar to systemic diffuse large B-cell lymphoma (sDLBCL), gene expression profiling in PCLBCL has provided a molecular basis for the subdivision into this biologically distinct subgroup (2). Hierarchical clustering of PCLBCL, leg-type and PCFCL based on B-cell signatures demonstrated that PCLBCL-LT shows expression profiles resembling that of activated B cells. In sDLBCL, these expression patterns, indicative of different stages of B-cell differentiation, have strong prognostic significance (3, 4).

Besides molecular profiling of the tumour cells themselves, their interaction with the tumour microenvironment is currently the focus of interest in lymphoma research (5–7). In this context, Lenz et al. (8) have reported that the gene expression profile of the non-tumour cells significantly predicts biological behaviour, treatment response and survival in patients with sDLBCL. The so-called stromal-1 signature is related to extracellular matrix deposition and histiocytic infiltration. This genetic pattern portends a good prognosis. On the contrary, the denoted stromal-2 signature is associated with a poor prognosis. This signature comprises an overexpression of genes encoding proteins such as platelet endothelial cell adhesion molecule-1 (PECAM-1/CD31) or the receptor for vascular endothelial growth factor (VEGFR-2). Hence, it primarily reflects angiogenesis. Accordingly, high intratumoral microvessel density (MVD) (9) as well as overexpression of vascular endothelial growth factor (VEGF) and other proangiogenic cytokines are assigned a major role for the tumour biology of different types of systemic lymphoma (10–12). As a consequence, therapies targeting the microenvironment have gained increasing attention and tumour angiogenesis has evolved as an attractive molecular target for novel therapeutic strategies (6).

The angiogenic switch is essential for a continuous tumour growth beyond a certain threshold (13). The complex process of tumour neoangiogenesis is governed by a plethora of countervailing humoral and cellular factors, which either induce or repress angiogenesis (14). The multifunctional cytokine vascular endothelial growth factor VEGF-A (commonly referred to as VEGF) represents the leading proangiogenic factor. It is expressed by tumour as well as stromal cells belonging to the tumour microenvironment. Of note, beyond its paracrine effects on endothelial cells VEGF may serve as an autocrine growth factor for the tumour cell itself (10).

In PCBCL the impact of the tumour microenvironment, especially of the tumour-associated microvasculature, has not been systematically analysed to date. Therefore, we used immunohistochemical analyses to evaluate whether subtypes of PCBCL can be distinguished with respect to microvascular density and expression of VEGF with its auto- and paracrine functions. Indeed, all of these parameters were elevated in the more aggressive lymphoma subtype PCLBCL-LT and indicated a worse prognosis for this entity. Factors associated with angiogenesis could therefore serve as novel molecular biomarkers in cutaneous B-cell lymphoma and may represent attractive therapeutic targets in the future.

MATERIALS AND METHODS

Patients

Paraffin-embedded tissue of skin of 12 patients with PCLBCL-LT, 15 patients with PCFCL, 17 patients with PCMZL, 9 patients with cutaneous B-cell rich pseudolymphoma (PL) (including lymphadenosis cutis benigna) and secondary cutaneous infiltrates of 13 patients with sDLBCL were obtained from the archives of the Department of Dermatology and the Institute of Pathology, University of Wuerzburg. Tissues were collected during a time period from 1999 to 2011. Patients alive provided informed consent prior to entering the study according to the guidelines of the institutional ethics committee. Patients with cutaneous B-cell lymphoma were staged according to the WHO/EORTC classification, those with systemic B-cell lymphoma according to the Ann Arbor system. The clinical characteristics of the lymphoma patients are summarized in Table I.

Immunohistochemical studies

Immunohistochemical analysis. Immunohistochemistry for CD31, VEGF, VEGFR-1 and VEGFR-2 was performed on 5- μ m tissue sections, which were placed on glass slides. For immunohistochemistry, tissue sections were deparaffinized in xylol and rehydrated in graded alcohol. For antigen retrieval, slides were overlaid with antigen retrieval solution (Dako, Hamburg, Germany) and incubated in saturated steam for 20 min. For staining, slides were incubated with a murine polyclonal anti-CD31/PECAM-1 antibody (Dako, Hamburg, Germany), which highlights both blood and lymphatic endothelial cells, rabbit polyclonal anti-VEGF antibody (VEGF (A-20), Santa Cruz, Heidelberg, Germany), which recognizes the VEGF_{165,121} and ₁₈₉ splice variants, rabbit polyclonal anti-VEGFR-1 antibody (Flt1, Spring Bioscience, Pleasanton, CA, USA) and rabbit polyclonal anti-VEGFR-2 antibody (Flk-1, Lab Vision, Thermo Science, Cheshire, UK).

Table I. Clinical patient characteristics

	PCLBCL-LT <i>n</i>	PCFCL <i>n</i>	PCMZL <i>n</i>	sDLBCL <i>n</i>
Patients	12	15	17	13
Mean age, years	70	55	48	65
Female	6	6	9	7
Male	6	9	8	6
Multi-ocular lesions at time of diagnosis	6	7	7	12
Solitary lesion at time of diagnosis	6	8	10	1
Best response CR after first-line therapy	6	10	11	5
Overall response CR	5	11	11	6
Mean progression-free survival, months	12	25	26	26
Mean overall survival, months	27	60	60	43

CR: complete remission; PCLBCL-LT: primary cutaneous large B-cell lymphoma, leg-type; PCFCL: primary cutaneous follicle centre lymphoma; PCMZL: primary cutaneous marginal zone lymphoma; sDLBCL: systemic diffuse large B-cell lymphoma.

For detection of VEGFR-1 and -2, Ultravision Quanto Detection System based on HRP-polymer (TL-125-QHL, Lab Vision, Thermo Science, Cheshire, UK) was used, for visualization of VEGF and CD31 the Dako Real Detection System based on HRP (K5003, Dako, Hamburg, Germany). Control antibody staining using polyclonal rabbit antiserum was performed accordingly for VEGF, VEGFR-1 and VEGFR-2 using appropriate protocols.

Evaluation of immunohistochemical staining. For evaluation of MVD, slides were scanned in the light microscope at $\times 40$ magnification and so-called hot spots were identified. MVD was determined by counting the number of CD31+ microvessels per measurement field at $\times 200$ magnification ($\times 20$ lens, $\times 10$ ocular, measurement field of 0.3136 mm²) using an Olympus BX45 microscope, as described (15). A microvessel was defined as any distinct CD31+ cell cluster with or without a vessel lumen. The cell surface molecule CD31, which is expressed both by blood as well as by lymphatic endothelium, was chosen for the immunohistochemical analyses because *CD31/PECAM-1* is one of the genes included in the formerly described stromal-2 signature (8). Counting was performed at least in 2 different representative fields of the tumour. The microvessel counts in the analysed areas were averaged and denoted MVD. To evaluate differences in microvasculature dependent on body location among patients with PCLBCL-LT, MVD inside the tumour and the non-involved surrounding tissue was assessed separately for location at the leg and non-leg location.

A visual semiquantitative scoring system was used for evaluating the staining results of VEGF and VEGFR-1 and -2. In case of VEGF, less than 5% cellular staining inside the tumour was scored negative (i.e. 0), 5–30% staining was scored weakly positive (i.e. 1), 30–60% staining intermediately positive (i.e. 2) and more than 60% staining strongly positive (i.e. 3). Due to lower frequencies of VEGFR-1- and VEGFR-2- positive cells, 5–20% of stained cells was graded as weakly (i.e. 1), 20–40% as intermediately (i.e. 2) and >40% staining as strongly positive (i.e. 3). Physiologically VEGF/VEGFR-positive structures, such as keratinocytes, endothelial cells or sweat glands, served as internal controls (16). Images of the immunohistological staining were acquired using a Zeiss Axiophot microscope and Axiocam digital camera.

Statistical analysis

Statistical analysis was performed with Prism Graph Version 3.0. Survival curves were estimated using log-rank test and

presented in Kaplan–Meier curves. Multivariate analysis was performed according to the Cox proportional hazards regression model. The χ^2 test for independence was used to test for correlations between categorical variables. Test results with p -values ≤ 0.05 were considered significant.

RESULTS

PCLBCL-LT exhibits a proangiogenic micromilieu

The patient characteristics are presented in Table I. Microvascular density was assessed to determine if tumour-associated vasculature correlates with the prognostic subtype of cutaneous B-cell lymphoma. All analysed tumours showed a significantly higher microvascular density than the surrounding normal tissue ($p \leq 0.001$). However, the tumour vascularity varied within and between the different subgroups (Fig. 1a). PCLBCL-LT (Fig. 1b) and sDLBCL displayed a significantly higher vascular density ($p \leq 0.001$) compared with PCFCL (Fig. 1c), PCMZL and PL. The score of intratumoral microvessel density was comparable among the subgroups PL, PCFCL and PCMZL. In accordance with high MVD PCLBCL-LT and sDLBCL showed significantly higher frequencies ($p \leq 0.0001$) of VEGF-positive tumour cells within the tumour than the other lymphoma subtypes and pseudolymphoma (Fig.

2 a, b). MVD both inside the tumour and in the surrounding tissue did not significantly differ for location at the leg and other locations in PCLBCL-LT (Fig. 3a). However, VEGF scores were higher in tumours and surrounding non-involved tissue when located in the lower extremities (Fig. 3 b, c).

Increased expression of VEGFR-1 and VEGFR-2 in tumour cells from PCLBCL-LT

In order to elucidate whether VEGF may potentially act on lymphoma cells in an autocrine manner, we analysed the expression pattern of VEGF receptors VEGFR-1 and VEGFR-2. Immunohistochemical scores for VEGFR-1 and VEGFR-2 expression were significantly higher in PCLBCL-LT and sDLBCL in comparison with PCFCL, PCMZL and PL ($p \leq 0.001$) (Fig. 2 c–f).

Microvessel density and prognosis in PCLBCL-LT

Overall survival (OS) and progression-free survival (PFS) of 12 patients with PCLBCL-LT being treated with R-CHOP \pm radiotherapy were correlated with MVD (Fig. 3 d, e). There was no statistically significant difference between the 2 groups, and confirmation in a larger cohort is required.

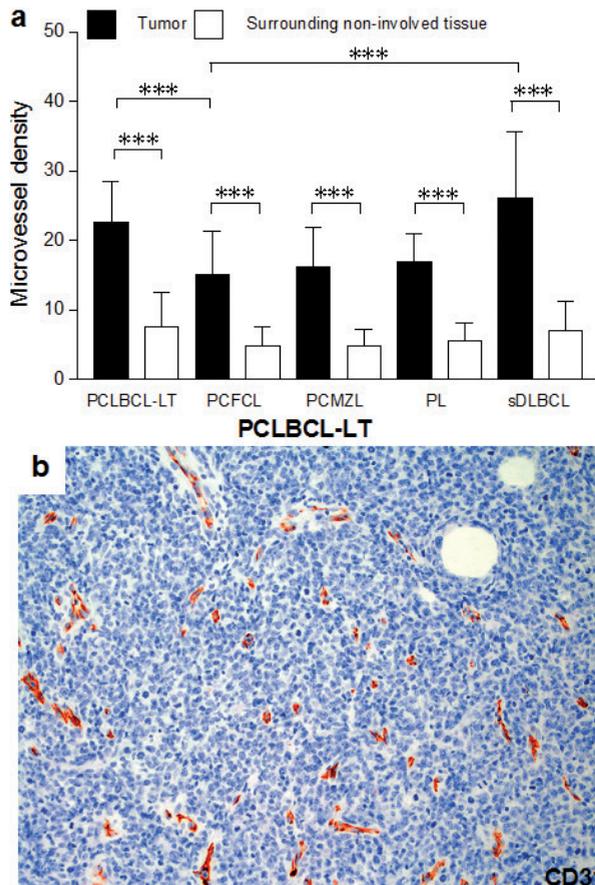


Fig. 1. (a) Microvascular density in the different subtypes of cutaneous B-cell lymphoma, B-cell pseudolymphoma and diffuse large B-cell lymphoma was determined by counting CD31 positive vascular structures. Vessel count/0.3136 mm² is separately indicated for the central part of the tumour (left column) and the surrounding non-involved tissue (right column). Primary cutaneous large B-cell lymphoma, leg-type (PCLBCL-LT) and systemic diffuse large B-cell lymphoma (sDLBCL) show significant higher microvessel density (MVD) than the indolent lymphoma subgroups. *** $p \leq 0.001$. (b) Example of CD31-staining demonstrating high density of tumour-associated blood vessels in PCLBCL-LT. Magnification $\times 200$. (c) Intermediate MVD in primary cutaneous follicle centre lymphoma (PCFCL) as assessed by immunohistochemistry with an antibody against CD31. Original magnification $\times 200$.

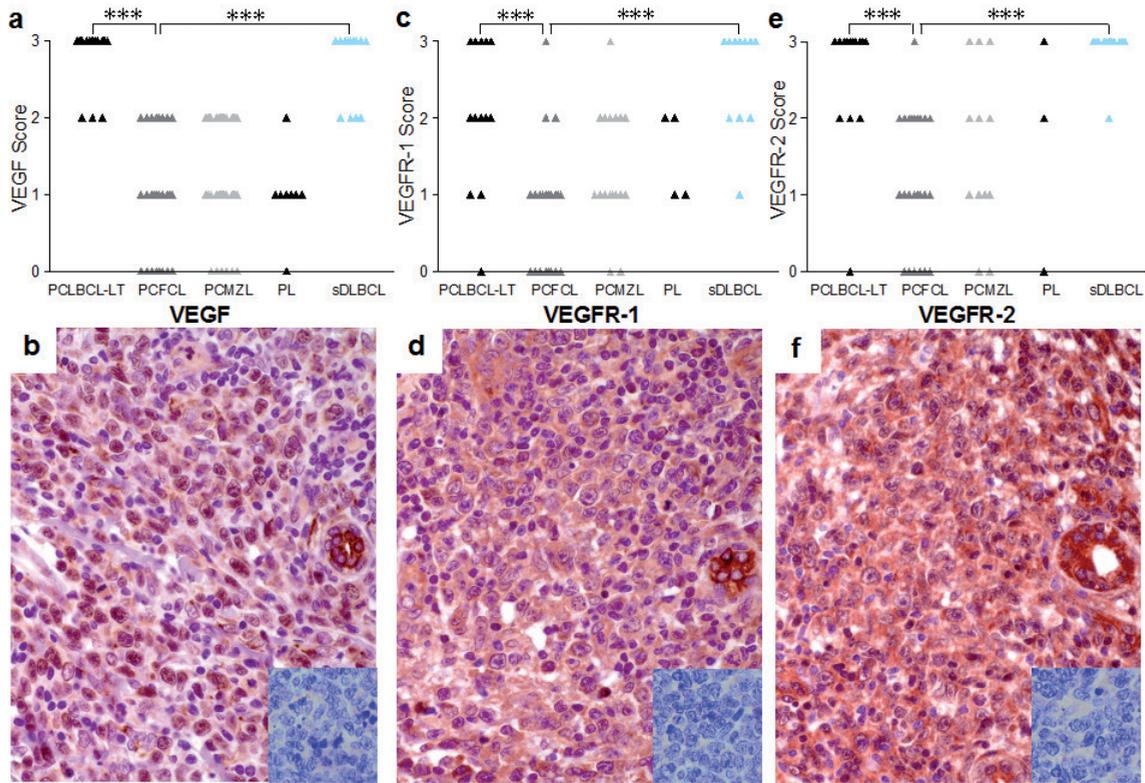


Fig. 2. Primary cutaneous large B-cell lymphoma, leg-type (PCLBCL-LT) and systemic diffuse large B-cell lymphoma (sDLBCL) exhibit significantly higher scores of (a) vascular endothelial growth factor (VEGF) and its receptors (c) VEGFR-1 and (d) VEGFR-2 than primary cutaneous follicle centre lymphoma (PCFCL), primary cutaneous marginal zone lymphoma (PCMZL) and pseudolymphoma (PL). $***p \leq 0.001$. Representative photomicrographs showing immunohistochemical stains for (b) VEGF, (c) VEGFR-1 and (e) VEGFR-2, in a representative specimen of the same patient with PCLBCL-LT. Original magnification $\times 400$. Control antibody staining (inset). Magnification $\times 400$.

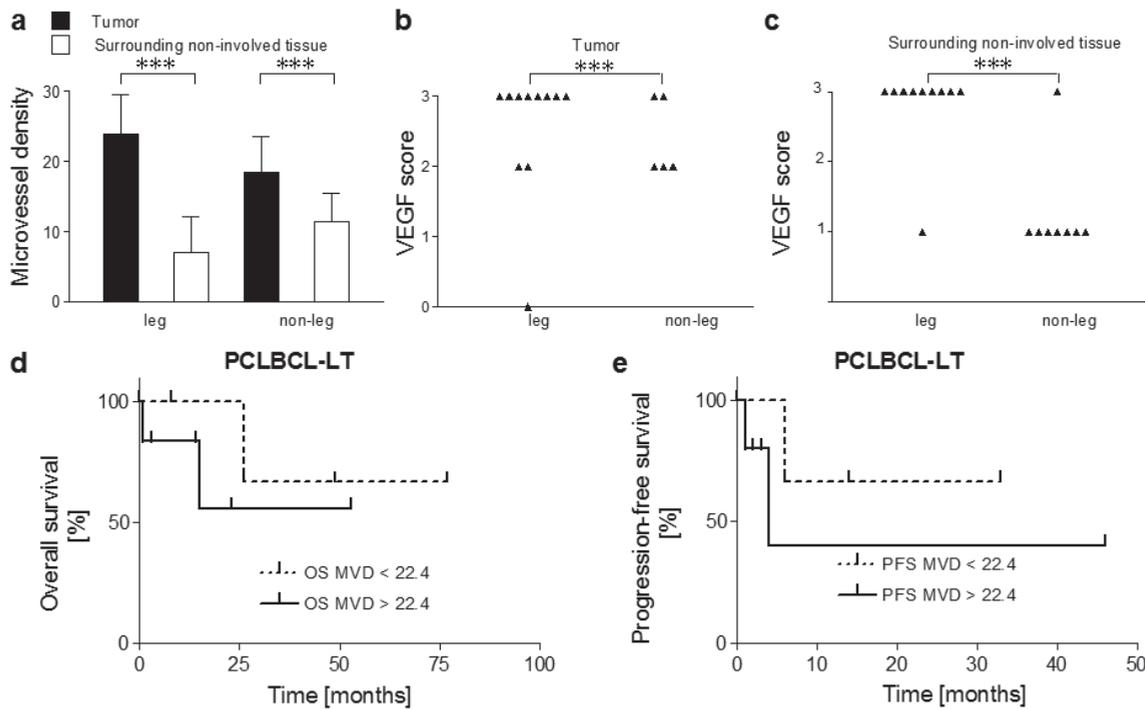


Fig. 3. Microvessel density (MVD) and expression of vascular endothelial growth factor (VEGF) expression was assessed dependent on body site. (a) MVD in primary cutaneous follicle centre lymphoma-leg-type (PCLBCL-LT) inside the tumour (black columns) and in the non-involved surrounding tissue (white columns) did not differ for location at the leg and non-leg location. VEGF score inside (b) the lymphoma infiltrate and (c) the surrounding tissue of PCLBCL-LT showed significantly higher values for location at the legs. No statistical difference is detected for low or high MVD on (d) overall and (e) progression-free survival in PCLBCL-LT.

DISCUSSION

In various systemic B-cell neoplasms increased tumour neoangiogenesis was found to be associated with a more aggressive behaviour and a poor prognosis (17). Recently, the underlying molecular basis has been delineated in sDLBCL (8). To date, no study has systematically addressed this topic in PCBCL. Therefore, we aimed to evaluate the role of tumour neoangiogenesis for the prognostically different subtypes PCFCL, PCMZL and PCLBCL-LT.

In the current study, all analysed specimens, both malignant lymphomas and benign pseudolymphomas, showed a significantly higher mean MVD compared with the surrounding normal tissue. Physiological angiogenesis is known from benign conditions, such as wound healing, hair growth cycle, acute and chronic inflammation (18). Likewise, it is herein observed in reactive inflammatory pseudolymphoma. Inflammatory processes induce vessel dilatation and enhanced permeability. Angiogenic response to inflammation promotes a reversible enlargement of pre-existing vessels (19) rather than sprouting of new vessels.

On the other hand, new vessel formation is induced via aberrant and constant overexpression of proangiogenic cytokines due to oncogene-driven gene expression. In our study, the microvessel count did not discriminate between benign inflammatory processes, such as PL and the indolent cutaneous lymphoma subtypes PCFCL and PCMZL. Only one prior study has to date assessed MVD in different types of indolent primary cutaneous B-cell lymphoproliferative disorders with comparable results (20). However, concerning the more aggressive lymphoma subtypes characterized by a large cell morphology (PCLBCL-LT, sDLBCL), significantly higher levels of mean MVD were detected. This is in line with previous studies in systemic B-NHL, where more unfavourable lymphoma subtypes showed higher MVD (9, 21). In systemic B-cell lymphomas the extent of MVD distinguishes between reactive benign lymphadenopathy and lymph nodes enlarged due to infiltration by non-Hodgkin's lymphoma (22). Hence, the assessment of MVD can serve as an important adjunct in the diagnostic algorithm for the diagnosis of large B-cell lymphoma of the skin (23).

To elucidate the pathophysiological background of neoangiogenesis in PCBCL, we analysed the protein expression of VEGF. As VEGF-A represents the leading proangiogenic mediator for regulating the complex process of neoangiogenesis (24), we restricted our analysis to this major cytokine. No difference in VEGF expression was observed in PL and indolent PCFCL and PCMZL corresponding to equal MVD in these subtypes. These findings may indicate that a proangiogenic milieu probably does not gain relevance until advanced stages of malignant transformation. In PCLBCL-LT

and sDLBCL, VEGF was significantly more strongly expressed, which corresponded to a higher vascularity. Similar studies in systemic haematological disorders have demonstrated higher levels of VEGF expression in tissue and circulation in more aggressive lymphoma subtypes (21, 25). To date, only 2 case reports have demonstrated elevated levels of VEGF in tissue and blood of a single patient with PCLBCL-LT (26, 27).

The exact nature of VEGF overexpression in PCLBCL-LT is so far unknown. One may speculate that higher VEGF expression in tumour cells of this lymphoma subtype may result from aberrant oncogenic signalling (28). PCLBCL-LT exhibits, among others, an upregulation of *bcl-2* and *c-myc* (2, 29). Beside anti-apoptotic and prosurvival properties (30), these factors are known to promote tumour angiogenesis (31–35). Hence, overexpression of *bcl-2*, which was detected in >90% of our PCLBCL-LT specimens (data not shown) and upregulation of *c-myc* may thus, at least partially, explain the observed VEGF induction in tumour cells of PCLBCL-LT. Of note, in our study VEGF expression was not only restricted to lymphoma cells. In addition, various stromal cells stained positive. This underlines the importance of the tumour microenvironment for the angiogenic response (36).

Induction of VEGF gene expression in response to hypoxia (37) strongly depends on the proliferative activity of the tumour (38). In this context, high expression of hypoxia inducible factors (*HIF-1 α* , *HIF-1 β*) and consecutive VEGF upregulation (39, 40) was demonstrated in aggressive lymphomas (41, 42). Intratumoral hypoxic conditions are supposed to be especially relevant for PCLBCL-LT. Its rapidly growing, ulcerating and necrotic tumours predominantly arise at body sites with restricted blood and oxygen supply, such as the lower legs (43). In chronic venous insufficiency, lipodermatosclerosis and peripheral arterial ischaemia of the lower extremities upregulation of VEGF was observed in response to hypoxia and inflammation (44–46). Likewise, in our study, higher amount of VEGF expression was observed in different specimen derived from the lower extremities. Hence, overexpression of VEGF in soft tissue of the lower extremities may establish a protumorigenic micromilieu and provides a rationale explanation for the prognostically unfavourable location at the legs in PCLBCL-LT.

To elucidate whether VEGF may exert beyond its paracrine, i.e. proangiogenic, also autocrine, i.e. protumorigenic, properties in PCBCL (10), we analysed the expression of VEGFR-1 and -2. In skin, VEGFR-1 and -2 are physiologically expressed on endothelial cells, adnexal structures and keratinocytes (16). Both receptors were also aberrantly upregulated on the tumour cells. Of interest, expression of both tyrosine kinase receptors was significantly higher in PCLBCL-LT and sDLBCL than in the indolent lymphoma subtypes and PL. A

functional autocrine loop with constitutive mitogenic self-stimulation in these more aggressive lymphomas may be the consequence. That could contribute to the high proliferation index, as observed in PCLBLT; accordingly, all analysed PCLBCL-LT showed a high proliferation index, as determined by Mib1 staining (data not shown).

PCLBCL-LT with higher mean baseline MVD (> 22.4 CD31⁺ microvessels/ 0.3136 mm^2) exhibited a slightly worse prognosis; however, without statistically significant difference. Traditional prognostic parameters for PCLBCL-LT include age, extent and location of skin lesions and expression of bcl-2, MUM-1 (29, 47). Further studies will clarify whether the easily-applicable assessment of VEGF- and VEGFR-expression as well as MVD by immunohistochemical staining with CD31 may provide a useful tool to more accurately predict the prognosis of PCBCL and to deduce the adequate therapeutic regimen. In systemic large B-cell lymphoma and other lymphoma subtypes antiangiogenic therapeutic modalities are already applied in clinical routine, and clinical studies are currently evaluating antiangiogenic therapies, including bevacizumab, endostar and thalidomide (www.clinicaltrials.gov).

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REFERENCES

1. Willemze R, Jaffe ES, Burg G, Cerroni L, Berti E, Swerdlow SH, et al. WHO-EORTC classification for cutaneous lymphomas. *Blood* 2005; 105: 3768–3785.
2. Hoefnagel JJ, Dijkman R, Basso K, Jansen PM, Hallermann C, Willemze R, et al. Distinct types of primary cutaneous large B-cell lymphoma identified by gene expression profiling. *Blood* 2005; 105: 3671–3678.
3. Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* 2000; 403: 503–511.
4. Bea S, Zettl A, Wright G, Salaverria I, Jehn P, Moreno V, et al. Diffuse large B-cell lymphoma subgroups have distinct genetic profiles that influence tumor biology and improve gene-expression-based survival prediction. *Blood* 2005; 106: 3183–3190.
5. Alizadeh AA, Gentles AJ, Alencar AJ, Liu CL, Kohrt HE, Houot R, et al. Prediction of survival in diffuse large B-cell lymphoma based on the expression of 2 genes reflecting tumor and microenvironment. *Blood* 2011; 118: 1350–1358.
6. Burger JA, Ghia P, Rosenwald A, Caligaris-Cappio F. The microenvironment in mature B-cell malignancies: a target for new treatment strategies. *Blood* 2009; 114: 3367–3375.
7. Eng C. Microenvironmental protection in diffuse large-B-cell lymphoma. *N Engl J Med* 2008; 359: 2379–2381.
8. Lenz G, Wright G, Dave SS, Xiao W, Powell J, Zhao H, et al. Stromal gene signatures in large-B-cell lymphomas. *N Engl J Med* 2008; 359: 2313–2323.
9. Cardesa-Salzman TM, Colomo L, Gutierrez G, Chan WC, Weisenburger D, Climent F, et al. High microvessel density determines a poor outcome in patients with diffuse large B-cell lymphoma treated with rituximab plus chemotherapy. *Haematologica* 2011; 96: 996–1001.
10. Gratzianger D, Zhao S, Tibshirani RJ, Hsi ED, Hans CP, Pohlman B, et al. Prognostic significance of VEGF, VEGF receptors, and microvessel density in diffuse large B cell lymphoma treated with anthracycline-based chemotherapy. *Lab Invest* 2008; 88: 38–47.
11. Alshenawy HA. Prognostic significance of vascular endothelial growth factor, basic fibroblastic growth factor, and microvessel density and their relation to cell proliferation in B-cell non-Hodgkin's lymphoma. *Ann Diagn Pathol* 2010; 14: 321–327.
12. Taskinen M, Jantunen E, Kosma VM, Bono P, Karjalainen-Lindsberg ML, Leppa S. Prognostic impact of CD31-positive microvessel density in follicular lymphoma patients treated with immunochemotherapy. *Eur J Cancer* 2010; 46: 2506–2512.
13. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med* 1971; 285: 1182–1186.
14. Baeriswyl V, Christofori G. The angiogenic switch in carcinogenesis. *Semin Cancer Biol* 2009; 19: 329–337.
15. Weidner N. The importance of tumor angiogenesis: the evidence continues to grow. *Am J Clin Pathol* 2004; 122: 675–677.
16. Man XY, Yang XH, Cai SQ, Bu ZY, Wu XJ, Lu ZF, et al. Expression and localization of vascular endothelial growth factor and vascular endothelial growth factor receptor-2 in human epidermal appendages: a comparison study by immunofluorescence. *Clin Exp Dermatol* 2009; 34: 396–401.
17. Koster A, Raemaekers JM. Angiogenesis in malignant lymphoma. *Curr Opin Oncol* 2005; 17: 611–616.
18. Huggenberger R, Detmar M. The cutaneous vascular system in chronic skin inflammation. *J Invest Dermatol Symp Proc* 2011; 15: 24–32.
19. Halin C, Detmar M. Chapter 1. Inflammation, angiogenesis, and lymphangiogenesis. *Methods Enzymol* 2008; 445: 1–25.
20. Schaefer L, Schmid MH, Mueller B, Dummer RG, Burg G, Kempf W. Angiogenesis in cutaneous lymphoproliferative disorders: microvessel density discriminates between cutaneous B-cell lymphomas and B-cell pseudolymphomas. *Am J Dermatopathol* 2000; 22: 140–143.
21. Tzankov A, Heiss S, Ebner S, Sterlacci W, Schaefer G, Augustin F, et al. Angiogenesis in nodal B cell lymphomas: a high throughput study. *J Clin Pathol* 2007; 60: 476–482.
22. Ho CL, Sheu LF, Li CY. Immunohistochemical expression of angiogenic cytokines and their receptors in reactive benign lymph nodes and non-Hodgkin lymphoma. *Ann Diagn Pathol* 2003; 7: 1–8.
23. Fernandez-Flores A, Smucler-Simonovich A, Escalante F, Manjon JA. The differential diagnosis between primary cutaneous large B-cell lymphoma and cutaneous follicular lymphoma: prognostic and therapeutic implications. *Am J Dermatopathol* 2011; 33: 819–826.
24. Tammela T, Enholm B, Alitalo K, Paavonen K. The biology of vascular endothelial growth factors. *Cardiovasc Res* 2005; 65: 550–563.
25. Passam FH, Sfiridaki A, Pappa C, Kyriakou D, Petreli E, Roussou PA, et al. Angiogenesis-related growth factors and cytokines in the serum of patients with B non-Hodgkin lymphoma; relation to clinical features and response to treatment. *Int J Lab Hematol* 2008; 30: 17–25.
26. Nakayama S, Yokote T, Kobayashi K, Hirata Y, Akioka T, Miyoshi T, et al. Primary cutaneous diffuse large B-cell

- lymphoma, leg type, with expression of both vascular endothelial growth factor and its receptors. *Leuk Res* 2009; 33: e181–e183.
27. Nakayama S, Yokote T, Kobayashi K, Hirata Y, Akioka T, Miyoshi T, et al. Primary cutaneous diffuse large B-cell lymphoma, leg type, with features simulating POEMS syndrome. *Eur J Haematol* 2010; 84: 79–83.
 28. Kim MK, Suh C, Chi HS, Cho HS, Bae YK, Lee KH, et al. VEGFA and VEGFR2 genetic polymorphisms and survival in patients with diffuse large B cell lymphoma. *Cancer Sci* 2011; 10: 7006.
 29. Hallermann C, Niermann C, Fischer RJ, Schulze HJ. New prognostic relevant factors in primary cutaneous diffuse large B-cell lymphomas. *J Am Acad Dermatol* 2007; 56: 588–597.
 30. Grange F, Petrella T, Beylot-Barry M, Joly P, D’Incan M, Delaunay M, et al. Bcl-2 protein expression is the strongest independent prognostic factor of survival in primary cutaneous large B-cell lymphomas. *Blood* 2004; 103: 3662–3668.
 31. Trisciuglio D, Gabellini C, Desideri M, Ragazzoni Y, De Luca T, Ziparo E, et al. Involvement of BH4 domain of bcl-2 in the regulation of HIF-1-mediated VEGF expression in hypoxic tumor cells. *Cell Death Differ* 2011; 18: 1024–1035.
 32. Kim JW, Gao P, Liu YC, Semenza GL, Dang CV. Hypoxia-inducible factor 1 and dysregulated c-Myc cooperatively induce vascular endothelial growth factor and metabolic switches hexokinase 2 and pyruvate dehydrogenase kinase 1. *Mol Cell Biol* 2007; 27: 7381–7393.
 33. Shchors K, Shchors E, Rostker F, Lawlor ER, Brown-Swigart L, Evan GI. The Myc-dependent angiogenic switch in tumors is mediated by interleukin 1beta. *Genes Dev* 2006; 20: 2527–2538.
 34. Hallermann C, Kaune KM, Siebert R, Vermeer MH, Tensen CP, Willemze R, et al. Chromosomal aberration patterns differ in subtypes of primary cutaneous B cell lymphomas. *J Invest Dermatol* 2004; 122: 1495–1502.
 35. Hallermann C, Kaune KM, Gesk S, Martin-Subero JI, Gunawan B, Griesinger F, et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6, and MALT1 gene loci in primary cutaneous B-cell lymphomas. *J Invest Dermatol* 2004; 123: 213–219.
 36. Burke B, Tang N, Corke KP, Tazzyman D, Ameri K, Wells M, et al. Expression of HIF-1alpha by human macrophages: implications for the use of macrophages in hypoxia-regulated cancer gene therapy. *J Pathol* 2002; 196: 204–212.
 37. Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: role of the HIF system. *Nat Med* 2003; 9: 677–684.
 38. Mizukami Y, Fujiki K, Duerr EM, Gala M, Jo WS, Zhang X, et al. Hypoxic regulation of vascular endothelial growth factor through the induction of phosphatidylinositol 3-kinase/Rho/ROCK and c-Myc. *J Biol Chem* 2006; 281: 13957–13963.
 39. Tsuzuki Y, Fukumura D, Oosthuysen B, Koike C, Carmeliet P, Jain RK. Vascular endothelial growth factor (VEGF) modulation by targeting hypoxia-inducible factor-1alpha--> hypoxia response element--> VEGF cascade differentially regulates vascular response and growth rate in tumors. *Cancer Res* 2000; 60: 6248–6252.
 40. Mazure NM, Chen EY, Yeh P, Laderoute KR, Giaccia AJ. Oncogenic transformation and hypoxia synergistically act to modulate vascular endothelial growth factor expression. *Cancer Res* 1996; 56: 3436–3440.
 41. Stewart M, Talks K, Leek R, Turley H, Pezzella F, Harris A, et al. Expression of angiogenic factors and hypoxia inducible factors HIF 1, HIF 2 and CA IX in non-Hodgkin’s lymphoma. *Histopathology* 2002; 40: 253–260.
 42. Giatromanolaki A, Koukourakis MI, Pezzella F, Sivridis E, Turley H, Harris AL, et al. Phosphorylated VEGFR2/KDR receptors are widely expressed in B-cell non-Hodgkin’s lymphomas and correlate with hypoxia inducible factor activation. *Hematol Oncol* 2008; 26: 219–224.
 43. Grange F, Beylot-Barry M, Courville P, Maubec E, Bagot M, Vergier B, et al. Primary cutaneous diffuse large B-cell lymphoma, leg type: clinicopathologic features and prognostic analysis in 60 cases. *Arch Dermatol* 2007; 143: 1144–1150.
 44. Peschen M, Grenz H, Brand-Saberi B, Bunaes M, Simon JC, Schopf E, et al. Increased expression of platelet-derived growth factor receptor alpha and beta and vascular endothelial growth factor in the skin of patients with chronic venous insufficiency. *Arch Dermatol Res* 1998; 290: 291–297.
 45. Herouy Y, Kreis S, Mueller T, Duerk T, Martiny-Baron G, Reusch P, et al. Inhibition of angiogenesis in lipodermatosclerosis: implication for venous ulcer formation. *Int J Mol Med* 2009; 24: 645–651.
 46. Choksy S, Pockley AG, Wajeh YE, Chan P. VEGF and VEGF receptor expression in human chronic critical limb ischaemia. *Eur J Vasc Endovasc Surg* 2004; 28: 660–669.
 47. Senff NJ, Hoefnagel JJ, Jansen PM, Vermeer MH, van Baarlen J, Blokx WA, et al. Reclassification of 300 primary cutaneous B-cell lymphomas according to the new WHO-EORTC classification for cutaneous lymphomas: comparison with previous classifications and identification of prognostic markers. *J Clin Oncol* 2007; 25: 1581–1587.