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Citation: Stejskal K, Streinzer M, Dyer A, Paulus HF, Spaethe J (2015) Functional Significance of Labellum Pattern Variation in a Sexually Deceptive Orchid (Ophrys heldreichii): Evidence of Individual Signature Learning Effects. PLoS ONE 10(11): e0142971. doi:10.1371/journal.pone.0142971

Editor: Renee M. Borges, Indian Institute of Science, INDIA

Received: April 23, 2015

Accepted: October 29, 2015

Published: November 16, 2015

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The authors thank the Austrian Science Fund (FWF): P21521-B17 for financial support. AGD acknowledges the Australian Research Council DP0878968/DP0987989/DP130100015 for financial support.

Competing Interests: AGD is an Academic Editor for PLOS ONE. This does not alter the authors' adherence to PLOS ONE Editorial policies and criteria.

RESEARCH ARTICLE

Functional Significance of Labellum Pattern Variation in a Sexually Deceptive Orchid (*Ophrys heldreichii*): Evidence of Individual Signature Learning Effects

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Abstract

Mimicking female insects to attract male pollinators is an important strategy in sexually deceptive orchids of the genus Ophrys, and some species possess flowers with conspicuous labellum patterns. The function of the variation of the patterns remains unresolved, with suggestions that these enhance pollinator communication. We investigated the possible function of the labellum pattern in Ophrys heldreichii, an orchid species in which the conspicuous and complex labellum pattern contrasts with a dark background. The orchid is pollinated exclusively by males of the solitary bee, Eucera berlandi. Comparisons of labellum patterns revealed that patterns within inflorescences are more similar than those of other conspecific plants. Field observations showed that the males approach at a great speed and directly land on flowers, but after an unsuccessful copulation attempt, bees hover close and visually scan the labellum pattern for up to a minute. Learning experiments conducted with honeybees as an accessible model of bee vision demonstrated that labellum patterns of different plants can be reliably learnt; in contrast, patterns of flowers from the same inflorescence could not be discriminated. These results support the hypothesis that variable labellum patterns in O. heldreichii are involved in flower-pollinator communication which would likely help these plants to avoid geitonogamy.

Introduction

Pollination by sexual deception is a rare and remarkable strategy in angiosperms as it constitutes an example of extreme floral specialization [1]. The majority of sexually deceptive plants are orchid species, although some cases have been reported for other plant families [2–4]. Whilst on a global basis, sexual deception is a rare strategy for plant pollination, it is interesting



that almost all species of the Mediterranean orchid genus *Ophrys*, which comprises more than 250 species, achieve pollination by exploiting the mate-seeking behavior of insect males [5-8]. The flowers imitate olfactory, visual and tactile signals of receptive females to attract males and provoke them to land on the labellum. During the so called pseudocopulation, the deceived males try to copulate with the labellum and in this process the pollinia are attached onto the males' body. During a subsequent visit by a male insect to another conspecific flower, the pollinia can then come into contact with the stigma, facilitating pollination [4,7,9-15]. As the mating attempts are unsatisfactory for the bee, the male's interest to visit other flowers usually decreases after a few minutes [6,13,14,16-18].

In the mating behavior of hymenopterans, which are the most frequent pollinators in the genus *Ophrys* [6], olfaction is the key sensory modality involved in male-female communication [19]. In order to attract only conspecific males, the sex pheromones produced by females are highly specific [4,20,21]. Due to the high specificity ensured by the imitated sex pheromones, visual signals were assumed to only play a minor role in pollinator attraction, and thus most *Ophrys* species are found to be visually inconspicuous and mainly possess flowers with achromatic cues to attract potential pollinators [8,15,22]. Recently, however, it was shown that some *Ophrys* species enhance pollinator attraction by means of color signals [23–25], and in addition to chromatic cues, some *Ophrys* species possess bright and highly complex line patterns that contrast well to the dark labellum [22,26]. Currently, the role of such patterns in some *Ophrys* species is poorly understood.

Flower patterns are widespread in angiosperms and show high variation in form and function. Some markings raise the effectiveness of flower visitation by guiding the visitors to the often elusive nectaries [27–30] or increase the tendency of regular flower visits instead of nectar robbing [31]. Other markings imitate pollen, anthers or stamen to attract pollinators without exposing the real pollen to environmental conditions [32,33] or simulate higher amounts of pollen to increase attractiveness to pollinators [34]. Additionally, some patterns serve as enticement for mate-seeking male insects [3,35]. However, many flower patterns are small and delicate, and their functional significance remains unknown. Moreover, the spatial resolving power of insect pollinators' compound eyes is typically considered to be relatively low [36], especially compared to the resolving power of a lens eye for viewing stimuli at a distance [37]. Thus it is not clear in many cases how and at what distance flower patterns are perceived by the pollinators [38].

In Ophrys, labella of the O. oestrifera and O. heldreichii group possess bright and highly complex patterns on a dark background [22]. It was supposed that these patterns mimic the wings or body-markings of the pollinator's females [6] but recently, Streinzer et al. [26] showed that during their approach to O. heldreichii flowers, the pollinators, males of the long-horned bee Eucera berlandi, do not discriminate between flowers with or without a pattern on their labellum as long as the olfactory signal is present [26]. An alternative hypothesis suggests that the labellum pattern might be learned by the unsatisfied males to avoid re-visitations [14]. As a consequence, to ensure subsequent visits by the males and thus successful pollen transfer, individual plants within a population are selected to possess distinct patterns to evade the association built between the negative experience of a male after an unsuccessful copulation and the corresponding pattern. This idea is supported by the observation that the labellum pattern of flowers from different plants appears distinct, whereas patterns from flowers of a single inflorescence have been reported to be more similar [14]. Recent work also shows that the sensory processing capabilities of bees have the capacity to use negative associations to promote high levels of perceptual learning within individual subjects, depending upon experience with visual stimuli and potential allocation of selective attention [39]. However, currently we are not aware of any systematic investigation and quantification of pattern similarity/dissimilarity within and



between inflorescences in the *O. oestrifera* and *O. heldreichii* group, nor if the visual processing of insects like bees is in principle able to discriminate between natural complex labellum patterns.

Our study thus aims to quantify the variation of labellum patterns within and between populations of *O. heldreichii* and test if bees can learn the patterns during their flower visit. In particular, we address the following questions:

- 1. How similar/different are labellum patterns among flowers of the same plant, among different plants within a population, and among different populations? We expect a higher similarity of patterns among flowers of the same plant than among different plants. We measured degree of pattern overlap, degree of symmetry, pattern surface area and contour density (defined as the ratio between contour length and the area of the pattern).
- 2. Do males attracted by a flower pay attention to the labellum pattern? We anticipate that the males attempt to learn and memorise the pattern after negative, non-rewarding encounters. We measured the time a male hovers in front of a flower before and after a copulation event, and calculated the mean distance to the labellum to estimate spatial resolution.
- 3. Are bees able to discriminate and learn patterns from flowers of either the same or different conspecific plants? We hypothesize that bees are able to discriminate patterns from flowers of different plants but fail to learn patterns of flowers of the same plant.

To answer these questions we observed and filmed behavioural sequences during pseudocopulation of *Eucera berlandi* male bees interacting with *Ophrys* flowers. In addition, we tested honeybees as an accessible classic model of insect and pollinator perceptual learning using appetitive-aversive differential conditioning experiments.

Materials and Methods

Study species

The genus *Ophrys* comprises c. 250 species and has its center of distribution in the Mediterranean region. Heldreich's bee orchid, *Ophrys heldreichii* SCHLECHTER, is widespread and abundant on Crete and possesses a labellum pattern which is characteristic for the *O. heldreichii* group [22], and was thus used in this study. Its pollinators are males of the long-horned bee *Eucera (Synhalonia) berlandi* DUSMET (Apoidea, Apidae, Eucerini) [6,25].

No specific permit was required since the locations of our field study were publicly accessible and only non-invasive techniques (filming and photographing) were applied.

Pattern analysis for quantification of similarity

To check if patterns of flowers from the same inflorescence are more similar than those of various plants, three populations of *O. heldreichii*, 15–17 plants per population, were investigated. Each of the inflorescences possessed 3–4 flowers, resulting in a total of 161 patterns suitable for analysis. Photographs of the labellum patterns were taken with a digital camera (Nikon D70s, Chiyoda, Japan) and modified in Adobe Photoshop CS4 by extending respective images to equal size (Fig 1A). The shape of each pattern was traced, and converted to an oval black and white picture using CorelDraw(R) Graphic Suite X3 and Image J 1.44 (Fig 1A). To quantify pattern similarity, patterns were randomly arranged in pairs and the following features were measured and compared: degree of pattern overlap, degree of vertical symmetry, relative pattern surface area and contour density. The degree of pattern overlap was measured by calculating the percentage of pixels, which were identical at the same position in a pair of patterns. A value close to one indicates a high match of the compared patterns, whilst a value approaching



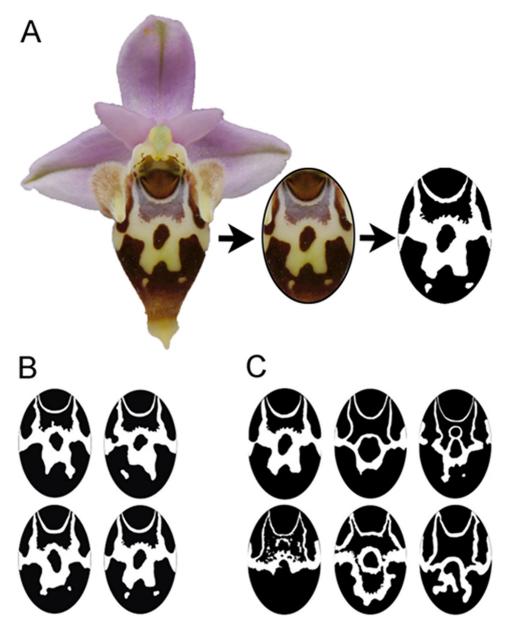


Fig 1. Labellum patterns of *O. heldreichii* **flowers.** (A) Transformation of a flower pattern to a black and white replica in an elliptical form used for discrimination experiments with honeybees. Patterns from flowers of the same inflorescence (B) appear more similar to each other than patters from flowers of different plant individuals (C).

zero indicates the patters were dissimilar. The similarity in degree of vertical symmetry was determined by drawing a vertical line in the center of the pattern and comparing each pixel on one side with the mirrored pixel on the opposite side to calculate the sum of deviation. A small value complies with a high degree of vertical symmetry. The relative pattern surface area was calculated as the ratio of the total number of black and white pixels. Finally, patterns were compared in terms of their contour density, measured as the ratio of pattern edge length and pattern area. For both relative pattern surface area and contour density, a high value indicates a



high match of the compared patterns. All measures were done in Adobe Photoshop CS4 Extended.

Scanning behavior

To analyze the pollinator's behavior during pseudocopulation, 10 males of *Eucera berlandi* approaching an *O. heldreichii* flower were filmed in the field (near Neapolis, Crete, Greece, 35° 15'N, 25°38'E) from above with a digital video camera (Sony DCR-SR50, Tokyo, Japan) at a rate of 25 frames s⁻¹. The filmed area was 26 x 15cm and a 10 x 10mm grid on the ground was used as reference [25] for reconstructing the flight paths of individual males. The videos were analyzed frame by frame with the tracking software SkillSpector 1.3.0 (Video4coach, Svendborg, Denmark). From these data we measured the flight duration before landing, the length of stay on the labellum, and the time the male hovered in front of the flower before leaving. In addition, we calculated the scanning duration at different distances from the labellum and the angular deviation of the bees' longitudinal body axis and the straight line between bee and flower.

Ophrys pattern discrimination experiments with honeybees

We aimed to understand how the patterns of *O. heldreichii* are involved in the recognition and identification of individual flowers by the pollinator. One important prerequisite is that the bees are capable of perceiving the patterns, or, more specifically, the difference between patterns, despite the relatively low spatial resolving power of their compound eyes.

Since *E. berlandi* males are typically rare in the field and fly only for few weeks during mating season, we conducted the learning experiments with honeybees as an accessible and classic model of insect visual perception [$\underline{40,41}$]. All bees possess apposition eyes and have comparable visual systems [$\underline{42}$]. A honeybee workers eye consists of 5735 ± 143 ommatidia [$\underline{43}$], while *E. berlandi* males possess 8354 ± 237 per eye (mean \pm SD; N = 5; Streinzer unpublished). The higher ommatidia number suggests a slightly higher visual acuity compared to honeybee workers (Land, 1997). Nevertheless, we anticipated useful information about the pollinator's pattern discrimination abilities from our behavioral experiments. For a detailed discussion about the limitations of transferring conclusions from our experiments with honeybee workers to *E. berlandi* males see discussion below.

The standardized black-and-white patterns (see above) were chosen for the learning experiment. The patterns were printed on light grey paper cards and laminated in mat laminating pouches (54 x 86mm) to both enable washing (to remove potential scents), and to eliminate specular reflections. Four patterns of flowers from the same inflorescence (Fig 1B) and six patterns of flowers from different plants (Fig 1C) were randomly selected. For each of the treatment groups, one pattern was chosen as a target, whereas the remaining ones served as distractors. One of the distractor patterns was further randomly selected and kept for the final transfer test (see below). Two experimental groups were tested, since patterns of different plants and patterns of the same plant respectively were presented.

The experiments were carried out with 16 honeybees (8 bees per treatment group), which is consistent with bee psychophysics studies [44–49]. Bees were trained individually from a feeding dish with 0.1M sugar solution to visit a vertical rotating screen of 60cm diameter, where 1M sugar solution was offered for correct landings on freely rotating hangers with a landing platform [44,45,50]. This experimental set-up prevented position learning and allowed the bees to choose any visual angle in order to recognize the stimuli [51].

During pre-training only one rewarding target was presented on the screen for 8 visits, which is a form of absolute conditioning [52]. For each landing the bee was allowed to drink a



10µl drop of sugar solution on the landing platform and another drop on a plastic stick, presented right beside the platform. While the bee was moved away from the screen sitting on the stick, the position of the target was changed by rotating the screen. To prevent any impact of olfactory cues, the landing platforms were covered with a transparent film, which was renewed after each foraging bout when the bee returned to the hive.

Following pre-training, each bee was trained with differential conditioning [52] with two rewarding targets and two unrewarding distractors presented on the screen for the next 140 decisions. To increase motivation, a landing on the (wrong) distractor stimulus was punished with a drop of bitter tasting quinine solution that honeybees are unable to detect with olfactory cues [39,53,54]. After training, a non-rewarded learning test that consisted of 20 decisions was conducted to collect data that completely excluded any possible use of olfactory or spatial cues. After refresher training, which means rewarding the bees to maintain their motivation, the bees were finally tested with a non-rewarded transfer test with two targets versus two distractors of a novel pattern for another 20 decisions.

During its approach to a pattern, a bee's response could be either correct (landing after approaching to the target stimulus or aborting an approach when approaching the distractor stimulus) or incorrect (landing on the distractor stimulus or aborting an approach to the target stimulus). A decision was considered as landing as soon as the bee was in contact with the stimuli or the landing platform. When the bee flew closer than 5cm to the stimuli and turned away afterwards without touching the stimuli or the landing platform, choices were counted as rejection [45].

To analyze such a complex decision making, signal detection theory has been proven useful in human psychology [55] but also to analyze behavior in bee learning experiments [45,56].

First, we calculated the probability of correct choices to the target stimulus (Pc, Eq 1) and incorrect choices to the distractor stimulus (Pi, Eq 2) for each training and test interval.

$$Pc = \sum \text{target landings} / (\sum \text{target landings} + \sum \text{target aborts})$$
 (Eq. 1)

$$Pi = \sum distractor\ landings / (\sum distractor\ landings + \sum distractor\ aborts)$$
 (Eq 2)

For each of these probabilities Z scores were calculated. The difference between the Z scores defines the variable d, where a d score of zero indicates chance performance and a value of 3.29 indicates perfect performance (for 10 decisions) [45]. To test, whether bee performance differed from chance, we calculated d scores for each individual and then tested the observed distribution of d scores against a value of zero using a one sample t-test. For convenience, we also present percent correct choices (target landings plus distractor aborts divided by total number of choices) as a more intuitive measure for performance. All statistical tests were performed on d scores.

Statistics

The statistical analyses were conducted using SPSS 15.0 (SPSS Inc.). To test if patterns of the same inflorescence are more similar than those of various plants a Mann-Whitney U-test was applied. For comparison between different populations a Kruskal-Wallis H-test was carried out. Differences between durations of approaching and scanning behavior were tested by a Mann Whitney U-test. To test whether honeybees are able to discriminate between flower patterns, we compared the d' scores for the last training block, the unrewarded test and the transfer test against an expected value of zero using a one-sample t-test (see above). All p-values above 0.05 were considered as statistically non-significant. When multiple comparisons were made, α -level was adjusted using Bonferroni correction.



Results

Pattern analysis for quantification of similarity

Labellum patterns of flowers from the same inflorescence showed a significantly higher degree of similarity (pattern overlap, degree of symmetry, relative pattern surface area, contour density) than patterns from different plants *within* each of the three populations (Fig 2). Interestingly, however, pattern similarity did not differ statistically *between* the populations, both for pattern similarity within the same plant (pattern overlap (H(2) = 4.155, p = 0.125), degree of symmetry (H(2) = 5.277, p = 0.071), relative pattern surface area (H(2) = 0.313, p = 0.855), contour density (H(2) = 2.439, p = 0.295) and between different plants (pattern overlap (H(2) = 1.869, p = 0.393), degree of symmetry (H(2) = 4.155, p = 0.125), relative pattern surface area (H(2) = 1.018, p = 0.601), contour density (H(2) = 0.0664, p = 0.967)).

Scanning behavior

Males approached the flower at high velocity (< 1s flight time in the observed area of 390cm²) and attempted to copulate with the labellum for 5.7s (median; min. 0.7s, max. 38.9s; Fig 3). After breaking off the copulation, males did not immediately leave the flower but hovered in

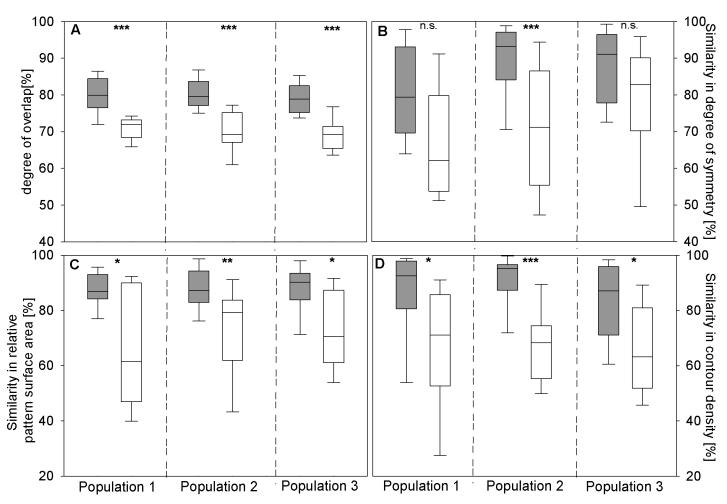


Fig 2. Degree of similarity among patterns from three *O. heldreichii* populations. Each box consists of 15 comparisons of randomly chosen flowers of the same (dark grey) or from different inflorescences (light grey). Four different features were compared: (A) pattern overlap, (B) pattern symmetry, (C) relative pattern surface area, and (D) contour density. Boxes represent inter-quartile range with median values. Stars indicate statistical differences after Bonferroni correction (***, p<0.00025; **, p<0.0125; n.s., not significant).

doi:10.1371/journal.pone.0142971.g002



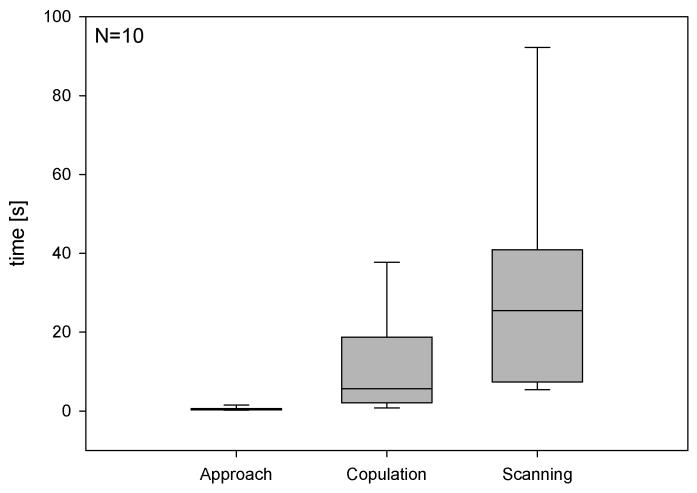


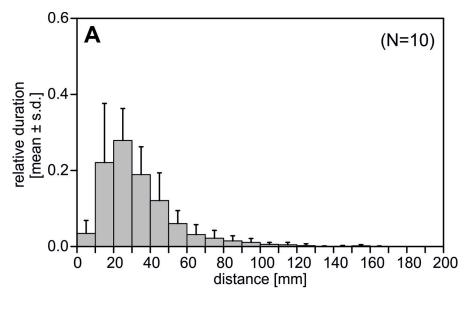
Fig 3. Duration of approach, copulation and post-copulation scanning behavior of *Eucera berlandi* males on an *Ophrys heldreichii* flower. Boxes represent inter-quartile range with median values, whiskers represent the 5 and 95 percentiles.

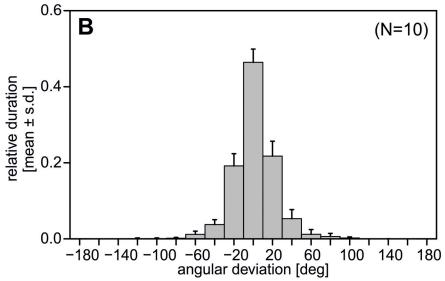
front of it, facing the labellum, for 25.4s (median; min. 5.7s, max. 97.2s; Figs $\underline{3}$ and $\underline{4C}$). The durations of approaching and post-copulation scanning behavior differed significantly (T = 55.0, p<0.001). During this "scanning behavior" they swung to the left and right while constantly facing the pattern head-on (Fig 4B) and kept an median flight distance of 2.0–4.1cm (range of medians; N = $10 \underline{\text{Fig 4A}}$) to the labellum, which corresponds to visual angles of 16° - 30° as viewed by the males (Fig 4).

Ophrys pattern discrimination experiments with honeybees

While patterns of flowers from different plants could be learnt and discriminated to a high level of accuracy consistently above 70% by one group of bees (d': 2.23±0.50; mean±s.d.; one sample t-test, t = 12.74, d.f. = 7, P<0.001; Fig 5), the group of bees trained with patterns of flowers from the same plant, were not able to distinguish between patterns of the 'same' inflorescence, even after a long training period of 140 decisions (d': 0.10±0.49; one sample t-test, t = 0.58, d.f. = 7, P = 0.58; Fig 5). The performance during the unrewarded test was significantly better than expected by random choice when patterns of flowers from different plants were presented (d': 2.28±0.31; one sample t-test, t = 20.54, d.f. = 7, P<0.001), but at chance level







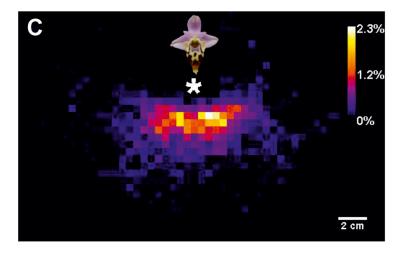




Fig 4. Scanning behavior of *Eucera berlandi* males after pseudocopulation on an *Ophrys heldreichii* flower (N = 10 flights). (A) Relative scanning duration at various distances from the flower. (B) Angular deviation of the bees' longitudinal body axis and the straight line between bee and orchid flower. (C) Bee position during a scanning flight in one representative trial in top view (sequence length 100 seconds). Colors represent the probability of presence in each 5x5mm pixel. The asterisk marks the position of the flower. Error bars show the standard deviation from the mean in (A) and (B).

when patterns of flowers from the same plant were presented (d': 0.32±0.39; one sample t-test, t = 2.30, d.f. = 7, P = 0.055; Fig 5).

The additional transfer tests revealed that bees did not only learn to discriminate between the training stimuli, but were able to choose the correct pattern in a novel stimuli combination when the distractor came from a novel plant (d': 1.82±0.43; one sample t-test, t = 12.05, d.f. = 7, P<0.001), but not when the distractor was a novel pattern from the same plant (d': 0.04±0.36; one sample t-test, t = 0.28, d.f. = 7, P = 0.79; Fig 5).

In both experiments it appears that bees chose more frequently a distractor stimulus than a rewarded target during the first training block (Fig 5). However, this is most likely due to the way decisions were scored rather than a preference for the distractor. Specifically; once a bee had chosen a *distractor* stimulus for the first time, she sometimes elected to land again on the platform, because she spent some time recognizing and becoming adjusted to the novel situation in the training procedure where now four stimuli were presented simultaneously (two targets and two distractors) in contrast to only one rewarded target (and no distractor) during pre-training. However, when she instead had chosen a *target* for the first time, she was immediately rewarded and removed from the disk by the plastic stick, which was counted as only one (correct) decision.

Discussion

In the present study we tested whether the variation in bright and complex patterns on the labellum of *O. heldreichii* flowers can act as a visual cue that could promote learning by a pollinator after pseudocopulation, and thus be potentially beneficial for both the pollinator and the plant.

The results are surprising in many aspects since classical models of insect visual perception only suggested that bees could process coarse elemental cues but were not capable of fine discriminations [57–59]. However, recent work from several groups has questioned such models of insect visual perception since bees can learn very complex visual stimuli like human faces [44], classic paintings [46] or pictures of landscapes [45,47], and other insects like the wasp *Polistes fuscatus* can reliably use vision to recognise the faces of conspecifics [60–62]. The reason for such contradictions has not been clear, and the new evidence we present shows a clear biological case where insects benefit from such impressive vision capabilities; which in some cases approaches the limit of what mammalian brains can achieve with visual processing [46,48,49]. In particular, we show that the signal provider, *O. heldreichii* may receive a benefit by promote allogamy, whilst the fine spatial vision of bees potentially provides individuals with a capacity to learn beneficial or non-beneficial stimuli through experience.

A variety of different theories have been developed to understand how insects like bees visually perceive the world [41,59,63,64]. Early theories were largely based upon the use of specific cues [59,65–67], but later work showed that bees can use both cues or even more sophisticated visual learning like configurations [48,68,69] and that these differences were due to both conditioning procedure and training length [52,70]. Indeed, recent work shows that attention is likely to be an important mechanism in bee choices since individual honeybees can learn complex hierarchical visual stimuli containing both local and global cues; and can modulate



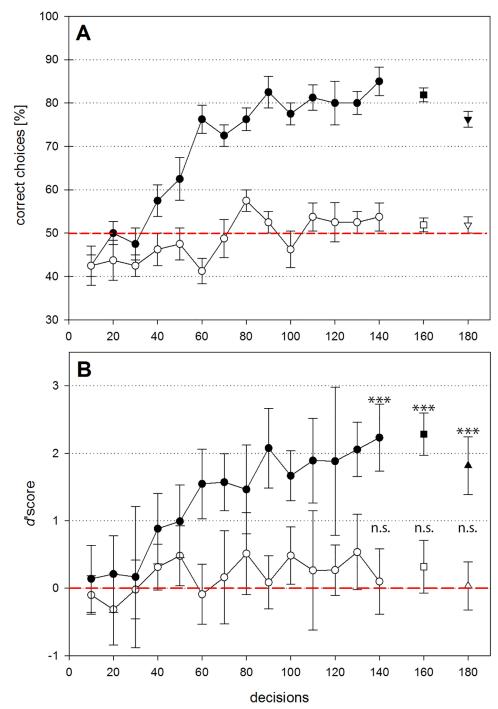


Fig 5. Honeybee performance during pattern discrimination experiments on a rotating screen. (A) Percent correct choices (target landings plus distractor aborts divided by total number of choices) during the pattern discrimination experiment. (B) Distribution of *d*'scores. Perfect discrimination would result in *d*'scores of 3.29, whereas values of zero indicate chance level performance. Filled circles show discrimination of pattern from different plants, open circles show discrimination of patterns from the same inflorescence. The square (after 160 decisions) symbolizes the performance in the unrewarded test, the triangle (after 180 decisions) the performance in the transfer test. Stars indicate statistical difference from chance (***p<0.001; n.s., not significant). Statistical tests were performed on *d*' scores only (see Results).



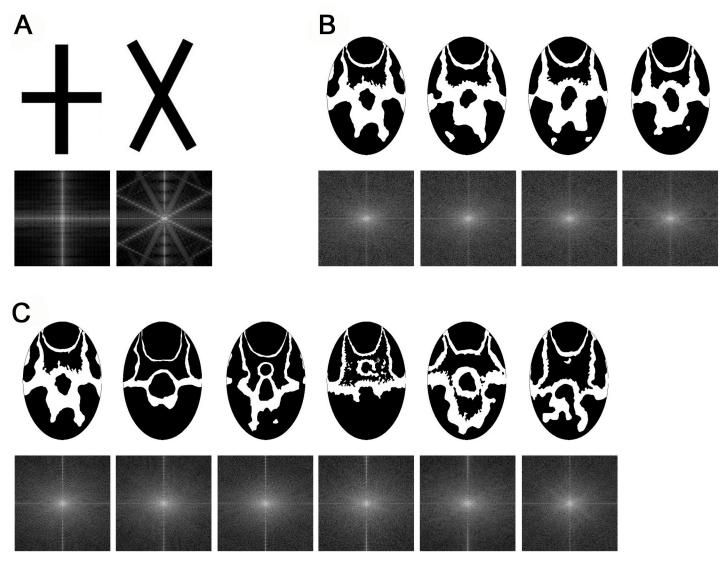


Fig 6. Fast Fourier Transform (FFT) of stimuli used in the honeybee experiments. Whereas the two types of crosses (A) differ widely in their FFT, the Ophrys pattern of same inflorescences (B) as well as those from different plants (C) show high similarity in their FFTs.

preferences towards either local or global cues depending upon experience or priming effects [71]. A comparison of labellum patterns in our experiments revealed a higher similarity within an inflorescence, whilst between inflorescences all tested cues revealed significantly higher variation (Fig 2). These potential floral cues are consistent with cues suspected to be involved in some forms of pattern recognition capability of bees [65,66,72]. Indeed the quantified higher similarity in these cues did result in more difficulties for honeybees learning the pattern discrimination tasks, suggesting our similarity specification is a valuable tool for evaluating how complex visual stimuli may be processed by bees. Another way that previous authors have quantified perceptual similarity of spatial stimuli is by measuring the Fast Fourier Transforms (FFT) of images [45–48]. We thus also measured the FFT of *Ophrys* patterns compared to rather simple 'cross' stimuli that differed greatly in spatial content of information. In comparison to the FFT for the cross stimuli, the various *Ophrys* labellum patterns show high level of



similarity in their FFTs (Fig 6), thus also supporting our metric of similarity using a method comparable to previous work.

Field observations of E. berlandi males approaching an O. heldreichii flower demonstrated that the pollinators, attracted by the scent, very quickly approached the flower without taking time to inspect the labellum pattern, suggesting that the pattern is not involved in male attraction [26]. However, once an individual bee was deceived by the plant, it hovered in front of the flower and appeared to inspect the detail of the labellum pattern (Fig 4D). This observed scanning behaviour was previously reported in several studies about shape discrimination experiments with honeybees [73,74] and provides support for previous assumptions that, following pseudocopulation, males try to memorize the plant's pattern and scent to prevent or reduce any further perceptual errors [11,13,14,20]. These previous findings are consistent with our current results of pattern discrimination experiments with honeybees using the rotating screen, which showed that bees were able to discriminate between patterns of different plant individuals, but failed to discriminate between patterns from flowers of the same inflorescence. The bees' performances in the unrewarded tests are in agreement with the performance of the last training block, thus allowing us to dissect that it is possible to make the discrimination solely on the basis of visual cues since the unrewarded tests were with fresh stimuli that exclude all olfactory cues. However, we have to act with caution when generalizing the results obtained with our honeybee model to other bee species, but we believe that our major conclusions hold also for E. berlandi males. Due to a slightly higher ommatidial number E. berlandi males should possess a slightly higher spatial resolving power [36] and thus should be able to discriminate fine patterns at least as well as honeybees do. Learning in honeybees was slow and it took at least 50 decisions to significantly discriminate the two patterns (Fig 5). In contrast, E. berlandi males usually visit orchids only a few times before they lose interest [75] and thus need to learn much faster. In addition, E. berlandi males are faced with aversive learning, whereas in honeybees we used appetitive conditioning. The learning speed and strength of an association is modulated by a number of factors. For example, the costs of a honeybee worker for choosing the wrong stimulus is low (few seconds of foraging time) since she can easily fly to the next stimulus. In contrast, competition among E. berlandi males for virgin females is extremely high due to high male numbers and few females which mate only once. Thus the decision to land on an orchid flower may prevent a male from detecting one of the rare females and might cause significant fitness consequences for the male. Our observations, that E. berlandi males spend a long time scanning, but honeybees made pretty fast decisions indicates different attentional levels and also supports this hypothesis. Due to these potentially high fitness costs learning speed is most likely higher in mate-seeking males than in bees foraging for food.

The learning and avoidance of individual flowers by the pollinator would lead to negative frequency-dependent selection, which may explain the high variation of labellum patterns among *O. heldreichii* flowers, since patterns, which are more similar to each other, are probably more often rejected in comparison to different ones [4,76]. As a consequence, flowers with rare patterns would receive more visits which probably leads to higher reproductive success. Previous work has documented a comparable variation in another modality, in particular odour signals of *Ophrys sphegodes*. Minimal differences in the odour components led to avoidance of previously visited flowers, but not of others [20]. The pollinator, *Andrena nigroaenea* males seem to reliably learn and memorize an individual flower odour bouquet and avoid future visits after an unsuccessful copulation event.

Recently, Sedeek et al. [77], investigated trait variation and reproductive isolation in four closely related species of *Ophrys*. They report a high intra-specific variation in labellum speculum shape and suggest, in line with our working hypothesis, that this variation may function in learning and subsequent avoidance of already visited flowers. However, no detailed



information is available with respect to the difference between inter-plant and between-plant speculum shape variation.

The higher similarity of labellum patterns within rather than among plants is consistent with selection, but also consistent with neutrality. It is likely that flowers of the same plant are more similar due to genetic and developmental constraints, as it had been shown for flower colour and pattern [78]. It would be important to examine in future studies if the pattern similarity with a plant in *O. heldreichii* is due to developmental constraints or indeed caused by selection by the pollinator [79,80]. In the latter scenario we predict that rare pattern types receive more pollinator visits and thus have a higher reproductive success than more common pattern types.

Aversive learning of individual flower traits may be an important strategy by sexually deceptive orchids in general, to reduce geitonogamy and thus promote allogamy. Recent investigations in the Australian sexually deceptive orchid genus Chiloglottis suggest that high levels of outcrossing may help to circumvent the costs, i.e. reduced seed germination and growth, of geitonogamous pollination [81]. Most species in the genus Ophrys possess inconspicuous labella without highly contrasting patterns. Therefore we hypothesize that the presence of these strong visual signals might be correlated to the importance of the visual system in the pollinator's mating behaviour, as was already assumed for the coloration of the sepals in some Ophrys species, including O. heldreichii [25]. Eucera males are highly visually guided during mate search and thus visual signature learning may be more important in this system compared with the olfactory signature learning that was shown for the less visually guided males of Andrena nigroaenea [20]. However, pattern variation is also found in *Ophrys* species with a less conspicuous labellar speculum [75]. This suggests that visual pattern learning may also contribute to individual signature learning in less visually guided bee species. It will be necessary to investigate further Ophrys species in future studies to examine if the presence of labellum pattern variation is correlated to pollinators with distinct visual systems.

Supporting Information

S1 Fig. Experimental setup for learning experiments with honeybees. Two target and two distractor stimuli were presented on hangers on a vertically rotation screen of 60cm diameter. The shown enlarged stimuli were used in a pre-experiment. (TIF)

S1 Table. Raw data. (XLSX)

Acknowledgments

We thank Martin Burd for helpful comments on an earlier version of the manuscript.

Author Contributions

Conceived and designed the experiments: KS MS AGD HP JS. Performed the experiments: KS MS JS. Analyzed the data: KS MS AGD JS. Wrote the paper: KS MS AGD HP JS.

References

 Vereecken NJ, Wilson CA, Hötling S, Schulz S, Banketov SA, Mardulyn P (2012) Pre-adaptations and the evolution of pollination by sexual deception: Cope's rule of specialization revisited. Proceedings of the Royal Society Biological Sciences Series B 279: 4786–4794.



- Ellis AG, Johnson SD (2010) Floral mimicry enhances pollen export: the evolution of pollination by sexual deceit outside of the Orchidaceae. The American Naturalist 176: E143–E151. doi: 10.1086/656487 PMID: 20843263
- Schiestl FP (2010) Pollination: Sexual mimicry abounds. Current Biology 20: R1020–R1022. doi: 10.1016/j.cub.2010.10.019 PMID: 21145017
- Schiestl FP (2005) On the success of a swindle: pollination by deception in orchids. Naturwissenschaften 92: 255–264. PMID: <u>15931514</u>
- Schiestl FP, Ayasse M, Paulus HF, Lofstedt C, Hansson BS, Ibarra F, et al. (1999) Orchid pollination by sexual swindle. Nature 399: 421–421.
- **6.** Paulus HF, Gack C (1990) Pollinators as prepollinating isolation factors: evolution and speciation in *Ophrys* (Orchidaceae). Israel Journal of Botany 39: 43–79.
- Pouyanne M (1917) La fécondation des Ophrys par les insectes. Bulletin de la Société d'Histoire Naturelle de l'Afrique du Nord 8: 6–7.
- Kullenberg B, Bergström G (1976) Hymenoptera Aculeata males as pollinators of Ophrys orchids. Zoologica Scripta 5: 13–23.
- Dafni A (1984) Mimicry and deception in pollination. Annual Review of Ecology and Systematics 15: 259–278.
- 10. Kullenberg B (1950) Investigations on the pollination of Ophrys species. Oikos 2: 1–19.
- Paulus HF (1988) Co-evolution und einseitige Anpassungen in Blüten-Bestäuber-Systemen: Bestäuber als Schrittmacher in der Blütenevolution. Verhandlungen der Deutschen Zoologischen Gesellschaft 81: 25–46.
- Paulus HF (1997) Signale in der Bestäuberanlockung: Weibchenimitation als Bestäubungsprinzip bei der mediterranen Orchideengattung Ophrys. Verhandlungen der Zoologisch-Botanischen Gesellschaft in Österreich 134: 133–176.
- Paulus HF (2005) Pollination biology of central European orchids (Bestäubungsbiologie heimischer Orchideen). Die Orchideen Deutschlands. Uhlstädt-Kirchhasel: Arbeitskreise heimische Orchideen. pp. 98–140.
- Paulus HF (2007) Wie Insekten-Männchen von Orchideenblüten getäuscht werden—Bestäubungstricks und Evolution in der mediterranen Ragwurzgattung Ophrys. Denisia 20: 255–294.
- Kullenberg B (1961) Studies in Ophrys pollination. Uppsala: Almquist & Wiksells Boktrykeri AB. 1–340
 p.
- 16. Nilsson AL (1992) Orchid pollination biology. Trends in Ecology & Evolution 7: 255–259.
- Paulus HF, Gack C (1983) Untersuchungen des Ophrys fusca-Formenkreises in Südspanien. Ein Beitrag zum Biospezieskonzept der Gattung Ophrys. Die Orchidee (Sonderheft): 65–72.
- 18. Paulus HF, Gack C, Maddocks R (1983) Beobachtungen und Experimente zum Pseudokopulationsverhalten an Ophrys (Orchidaceae): Das Lernverhalten von Eucera barbiventris (Apoidea, Anthophoridae) an Ophrys scolopax in Südspanien. Senghas K; Sundermann H. Probleme der Taxonomie und Vermehrung europäischer und mediterraner Orchideen. Die Orchidee (Sonderheft) 1983: Brücke Verlag Hildesheim. pp. 73–79.
- Ayasse M, Paxton RJ, Tengö J (2001) Mating behavior and chemical communication in the order Hymenoptera. Annual Review of Entomology 46: 31–78. PMID: 11112163
- Ayasse M, Schiestl FP, Paulus HF, Löfstedt C, Hansson B, Ibarra F, et al. (2000) Evolution of reproductive strageties in the sexually deceptive orchid Ophrys sphegodes: How does flower-specific variation of odor signals influence reproducitve success? Evolution 54: 1995–2006. PMID: 11209776
- Schiestl FP, Ayasse M (2002) Do changes in floral odor cause speciation in sexually deceptive orchids? Plant Systematics and Evolution 234: 111–119.
- 22. Delforge P (2006) Orchids of Europe, North Africa and the Middle East. Oregon, USA: Timber Press.
- 23. Spaethe J, Moser W, Paulus HF (2007) Increase of pollinator attraction by means of a visual signal in the sexually deceptive orchid, *Ophrys heldreichii* (Orchidaceae). Plant Systematics and Evolution 264: 31–40.
- Spaethe J, Streinzer M, Paulus HF (2010) Why sexually deceptive orchids have colored flowers. Communicative & Integrative Biology 3: 139–141.
- 25. Streinzer M, Paulus HF, Spaethe J (2009) Floral colour signal increases short-range detectability of a sexually deceptive orchid to its bee pollinator. Journal of Experimental Biology 212: 1365–1370. doi: 10.1242/jeb.027482 PMID: 19376957
- Streinzer M, Ellis T, Paulus HF, Spaethe J (2010) Visual discrimination between two sexually deceptive Ophrys species by a bee pollinator. Arthropod Plant Interactions 4: 141–148. PMID: <u>21516265</u>



- 27. Manning A (1956) The effect of honey-guides. Behaviour 9: 114-139.
- 28. Waser NM, Price MV (1985) The effect of nectar guides on pollinator preference: experimental studies with a montane herb. Oecologia 67: 121–126.
- Dafni A, Kevan PG (1996) Floral symmetry and nectar guides: ontogenetic constraints from floral development, colour pattern rules and functional significance. Botanical Journal of the Linnean Society 120: 371–377.
- Leonard AS, Papaj DR (2011) 'X' marks the spot: The possible benefits of nectar guides to bees and plants. Functional Ecology 25: 1293–1301.
- Leonard AS, Brent J, Papaj DR, Dornhaus A (2013) Floral nectar guide patterns discourage nectar robbing by bumble bees. PLoS ONE 8: e55914. doi: 10.1371/journal.pone.0055914 PMID: 23418475
- **32.** Lunau K (2000) The ecology and evolution of visual pollen signals. Plant Systematics and Evolution 222: 89–111.
- Osche G (1979) Zur Evolution optischer Signale bei Blütenpflanzen. Biologie in unserer Zeit 9: 161– 170.
- 34. Lunau K, Wacht S (1997) Signalfunktion von Pollen. Biologie in unserer Zeit 27: 169-181.
- Johnson SD, Dafni A (1998) Response of bee-flies to the shape and pattern of model flowers: implications for floral evolution in a Mediterranean herb. Functional Ecology 12: 289–297.
- 36. Land MF (1997) Visual acuity in insects. Annual Review of Entomology 42: 147–177. PMID: 15012311
- Land MF, Chittka L (2013) Vision. The insects: structure and function. 5th Edition ed. Simpson S.J.;
 Douglas A.E., Cambridge: Campridge University Press. pp. 708–737.
- Dafni A, Lehrer M, Kevan PG (1997) Spatial flower parameters and insect spatial vision. Biological Reviews 72: 239–282.
- Avarguès-Weber A, De Brito Sanchez MG, Giurfa M, Dyer AG (2010) Aversive reinforcement improves visual discrimination learning in free-flying honeybees. PLoS ONE 5: e15370. doi: 10.1371/journal. pone.0015370 PMID: 20976170
- Srinivasan MV (2010) Honey bees as a model for vision, perception, and cognition. Annual Review of Entomology 55: 267–284. doi: 10.1146/annurev.ento.010908.164537 PMID: 19728835
- 41. Avarguès-Weber A, Deisig N, Giurfa M (2011) Visual cognition in social insects. Annual Review of Entomology 56: 423–443. doi: 10.1146/annurev-ento-120709-144855 PMID: 20868283
- Briscoe AD, Chittka L (2001) The evolution of color vision in insects. Annual Review of Entomology 46: 471. PMID: 11112177
- Streinzer M, Brockmann A, Nagaraja N, Spaethe J (2013) Sex and caste-specific variation in compound eye morphology of five honeybee species. PLoS ONE 8: e57702. doi: 10.1371/journal.pone. 0057702 PMID: 23460896
- Dyer AG, Neumeyer C, Chittka L (2005) Honeybee (Apis mellifera) vision can discriminate between and recognise images of human faces. Journal of Experimental Biology 208: 4709. PMID: 16326952
- 45. Dyer AG, Rosa MGP, Reser DH (2008) Honeybees can recognise images of complex natural scenes for use as potential landmarks. Journal of Experimental Biology 211: 1180–1186. doi: 10.1242/jeb. 016683 PMID: 18375842
- **46.** Wu W, Moreno A, Tangen J, Reinhard J (2013) Honeybees can discriminate between Monet and Picasso paintings. Journal of Comparative Physiology A 199: 45–55.
- Zhang SW, Srinivasan MV, Zhu H, Wong J (2004) Grouping of visual objects by honeybees. Journal of Experimental Biology 207: 3289–3298. PMID: 15326205
- Avarguès-Weber A, Portelli G, Benard J, Dyer A, Giurfa M (2010) Configural processing enables discrimination and categorization of face-like stimuli in honeybees. Journal of Experimental Biology 213: 593–601. doi: 10.1242/jeb.039263 PMID: 20118310
- Avarguès-Weber A, Dyer AG, Combe M, Giurfa M (2012) Simultaneous mastering of two abstract concepts by the miniature brain of bees. Proceedings of the National Academy of Sciences 109: 7481–7486.
- 50. Morawetz L, Svoboda A, Spaethe J, Dyer AG (2013) Blue colour preference in honeybees distracts visual attention for learning closed shapes. Journal of Comparative Physiology A Sensory Neural and Behavioral Physiology 199: 817–827.
- **51.** Efler D, Ronacher B (2000) Evidence against a retinotopic-template matching in honeybees' pattern recognition. Vision Research 40: 3391–3403. PMID: 11058736
- Giurfa M, Hammer M, Stach S, Stollhoff N, Müller-Deisig N, Mizyrycki C (1999) Pattern learning by honeybees: conditioning procedure and recognition strategy. Animal behaviour 57: 315–324. PMID: 10049470



- Dyer AG, Neumeyer C (2005) Simultaneous and successive colour discrimination in the honeybee (Apis mellifera). Journal of Comparative Physiology A 191: 547–557.
- 54. Rodríguez-Gironés MA, Trillo A, Corcobado G (2013) Long term effects of aversive reinforcement on colour discrimination learning in free-flying bumblebees. PLoS ONE 8: e71551. doi: 10.1371/journal.pone.0071551 PMID: 23951186
- Collishaw SM, Hole GJ (2000) Featural and configurational processes in the recognition of faces of different familiarity. Perception 29: 893–909. PMID: <u>11145082</u>
- Lynn SK, Cnaani J, Papaj DR, Björklund M (2005) Peak shift discrimination learning as a mechanism of signal evolution. Evolution 59: 1300–1305. PMID: 16050106
- Horridge A (2000) Seven experiments on pattern vision of the honeybee, with a model. Vision Research 40: 2589–2603. PMID: 10958911
- Horridge A (2009) Generalization in visual recognition by the honeybee (*Apis mellifera*): A review and explanation. Journal of Insect Physiology 55: 499–511. PMID: 19418594
- 59. Horridge A (2009) What does an insect see? The Journal of Experimental Biology 212: 2721–2729. doi: 10.1242/jeb.030916 PMID: 19684204
- Tibbetts EA (2002) Visual signals of individual identity in the wasp *Polistes fuscatus*. Proceedings of the Royal Society of London Series B: Biological Sciences 269: 1423–1428. PMID: 12137570
- Sheehan MJ, Tibbetts EA (2011) Specialized face learning is associated with individual recognition in paper wasps. Science 334: 1272–1275. doi: 10.1126/science.1211334 PMID: 22144625
- 62. Tibbetts EA, Dyer AG (2013) Good with faces. Scientific American 309: 62-67. PMID: 24383366
- 63. Srinivasan MV (2011) Honeybees as a model for the study of visually guided flight, navigation, and biologically inspired robotics. Physiological Reviews 91: 413–460. doi: 10.1152/physrev.00005.2010 PMID: 21527730
- 64. Dyer AG (2012) The mysterious cognitive abilities of bees: why models of visual processing need to consider experience and individual differences in animal performance. Journal of Experimental Biology 215: 387–395. doi: 10.1242/jeb.038190 PMID: 22246247
- Wehner R (1981) Spatial vision in arthropods. In: Autrum H, editor. Handbook of Sensory Physiology. Berlin: Springer-Verlag. 287–616.
- Giurfa M, Eichmann B, Menzel R (1996) Symmetry perception in an insect. Nature 382: 458–461. PMID: 18610516
- Lehrer M (1999) Shape perception in the honeybee: symmetry as a global framework. International Journal of Plant Sciences 160: 51–65.
- Stach S, Benard J, Giurfa M (2004) Local-feature assembling in visual pattern recognition and generalization in honeybees. Nature 429: 758–761. PMID: 15201910
- Dyer AG, Griffiths DW (2012) Seeing near and seeing far: behavioural evidence for dual mechanisms of pattern vision in the honeybee (*Apis mellifera*). Journal of Experimental Biology 215: 397–404. doi: 10.1242/jeb.060954 PMID: 22246248
- **70.** Stach S, Giurfa M (2005) The influence of training length on generalization of visual feature assemblies in honeybees. Behavioural Brain Research 161: 8–17. PMID: 15904705
- Avarguès-Weber A, Dyer AG, Ferrah N, Giurfa M (2015) The forest or the trees: preference for global over local image processing is reversed by prior experience in honeybees. Proceedings of the Royal Society B 2015; 282(1799): 20142384
- Lehrer M (1999) Dorsoventral asymmetry of colour discrimination in bees. Journal of Comparative Physiology A 184: 195–206.
- Anderson AM (1979) Visual scanning in the honey bee. Journal of Comparative Physiology A 130: 173–182.
- Lehrer M, Wehner R, Srinivasan M (1985) Visual scanning behaviour in honeybees. Journal of Comparative Physiology A. Sensory Neural and Behavioral Physiology 157: 405–415.
- Rakosy D, Streinzer M, Paulus HF, Spaethe J (2012) Floral visual signal increases reproductive success in a sexually deceptive orchid. Arthropod-Plant Interactions 6: 671–681. PMID: 23750181
- Gigord LDB, Macnair MR, Smithson A (2001) Negative frequency-dependent selection maintains a dramatic flower color polymorphism in the rewardless orchid *Dactylorhiza sambucina* (L.) Soò. Proceedings of the National Academy of Sciences 98: 6253–6255.
- 77. Sedeek KEM, Scopece G, Staedler YM, Schönenberger J, Cozzolino S, Schiestl FP, et al. (2014) Genic rather than genome-wide differences between sexually deceptive *Ophrys* orchids with different pollinators. Molecular Ecology 23: 6192–6205. doi: 10.1111/mec.12992 PMID: 25370335



- Mol J, Grotewold E, Koes R (1998) How genes paint flowers and seeds. Trends in Plant Science 3:212–217.
- Hopkins R, Rausher MD (2011) Identification of two genes causing reinforcement in the Texas wildflower Phlox drummondii. Nature 469: 411–414. doi: 10.1038/nature09641 PMID: 21217687
- Hopkins R, Rausher MD (2012) Pollinator-mediated selection on flower color allele drives reinforcement. Science 335: 1090–1092. doi: 10.1126/science.1215198 PMID: 22300852
- 81. Whitehead MR, Linde CC, Peakall R (2015) Pollination by sexual deception promotes outcrossing and mate diversity in self-compatible clonal orchids. Journal of Evolutionary Biology 28:8: 1526–1541. doi: 10.1111/jeb.12673 PMID: 26079670