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Lifetime stress accelerates epigenetic aging in an urban, African American cohort: relevance of glucocorticoid signaling

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Abstract

Background: Chronic psychological stress is associated with accelerated aging and increased risk for aging-related diseases, but the underlying molecular mechanisms are unclear.

Results: We examined the effect of lifetime stressors on a DNA methylation-based age predictor, epigenetic clock. After controlling for blood cell-type composition and lifestyle parameters, cumulative lifetime stress, but not childhood maltreatment or current stress alone, predicted accelerated epigenetic aging in an urban, African American cohort (n = 392). This effect was primarily driven by personal life stressors, was more pronounced with advancing age, and was blunted in individuals with higher childhood abuse exposure. Hypothesizing that these epigenetic effects could be mediated by glucocorticoid signaling, we found that a high number (n = 85) of epigenetic clock CpG sites were located within glucocorticoid response elements. We further examined the functional effects of glucocorticoids on epigenetic clock CpGs in an independent sample with genome-wide DNA methylation (n = 124) and gene expression data (n = 297) before and after exposure to the glucocorticoid receptor agonist dexamethasone. Dexamethasone induced dynamic changes in methylation in 31.2 % (110/353) of these CpGs and transcription in 81.7 % (139/170) of genes neighboring epigenetic clock CpGs. Disease enrichment analysis of these dexamethasone-regulated genes showed enriched association for aging-related diseases, including coronary artery disease, arteriosclerosis, and leukemias.

Conclusions: Cumulative lifetime stress may accelerate epigenetic aging, an effect that could be driven by glucocorticoid-induced epigenetic changes. These findings contribute to our understanding of mechanisms linking chronic stress with accelerated aging and heightened disease risk.

Keywords: Aging, Aging-related disease, DNA methylation, Epigenetics, Gene expression, Glucocorticoids, Psychological stress

Background

The last decades have witnessed a dramatic increase in life expectancy. As a result, the number of older adults is predicted to more than double over the next two decades [1, 2]. While this increase in life expectancy is undoubtedly one of the biggest achievements of modern medicine, population aging also brings forth

an unprecedented increase in aging-related diseases, including cardiovascular disease, cancer, and dementia [3]. Given that these conditions are currently the leading causes of morbidity and mortality, it is imperative to gain insights into factors that impact healthy aging and contribute to aging-related diseases.

An important risk factor for accelerated aging and aging-related diseases is psychological stress. Although stressors are ubiquitous in nature and necessary for survival [4], excessive and chronic stress has been associated with accelerated cellular aging [5, 6] and increased

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risk for aging-related disease phenotypes, including cardiovascular disease, immune dysregulation, and late-life neuropsychiatric disorders [7–12]. Furthermore, stressors occurring during sensitive developmental periods, such as childhood maltreatment, have been linked with later development of aging-related diseases [13–15]. Lastly, stress-related psychiatric disorders, including major depression and post-traumatic stress disorder (PTSD), are themselves risk factors for such diseases [15, 16]. Despite these observations, the molecular mechanisms linking psychological stress with accelerated aging and aging-related diseases remain largely unknown.

One plausible mechanism that may mediate the adverse effects of stress on the aging process is epigenetic regulation. Long-term epigenetic changes can be induced by environmental stimuli, including psychological stressors, and can shape complex phenotypes [17]. The most studied epigenetic modification in this context is DNA methylation. Stressors can induce lasting changes in DNA methylation [18, 19], an effect that is in part mediated by the genomic effects of glucocorticoids, a primary molecular effector of the stress response [20]. Glucocorticoids exert actions in essentially every body organ via activation of the glucocorticoid receptor (GR), a transcription factor that regulates gene expression by the binding of its homodimer to glucocorticoid response elements (GREs) in regulatory regions of target genes [21]. Beyond regulating gene transcription, GRE binding can locally induce lasting changes in DNA methylation, a form of molecular memory that shapes subsequent responses to glucocorticoids and stressors [17, 18, 22–24]. Therefore, it is plausible that stress and glucocorticoid exposure throughout the lifetime could impact cellular aging via cumulative effects on aging-related DNA methylation sites.

Aging and aging-related diseases are associated with profound changes in DNA methylation [25–31]. Recognizing the importance of DNA methylation in the aging process has led to recent development of several DNA methylation-based predictors of aging [27, 32–34]. Among these, a composite predictor comprised of 353 Cytosine-phosphate-Guanosine sites (CpGs) across the genome ('epigenetic clock') was shown to strongly correlate with chronological age across multiple tissues in humans [27], suggesting its usefulness as a biomarker in aging-related research. Using this predictor, accelerated epigenetic aging (Δ -age), defined as the difference between DNA methylation-predicted age (DNAM-age) and chronological age, has been associated with aging-related and other phenotypes, including cancer, obesity, cytomegalovirus infection, Down's syndrome, PTSD, physical and cognitive decline, all-cause mortality, and the presence of higher self-control and lower socioeconomic status [27, 35–41]. However, no studies have

examined the relationship between this predictor and cumulative lifetime stress nor the potential molecular mechanisms underlying this relationship.

In the present study, we first show that cumulative lifetime stress, but not childhood or current stress alone, is associated with accelerated epigenetic aging in a cohort of highly traumatized African American individuals. Examining GR signaling as a potential mechanism underlying this effect, we identify that a high number of epigenetic clock CpGs are located within functional GREs and show dynamic methylation changes following GR activation by exposure to the GR agonist dexamethasone (DEX). Lastly, we show that genes neighboring these CpGs are dynamically regulated by DEX and that these DEX-regulated genes show enriched association for aging-related diseases. Taken together, our findings support a model of stress-induced acceleration of epigenetic aging, overall contributing to our understanding of mechanisms linking chronic stress with accelerated aging and heightened disease risk.

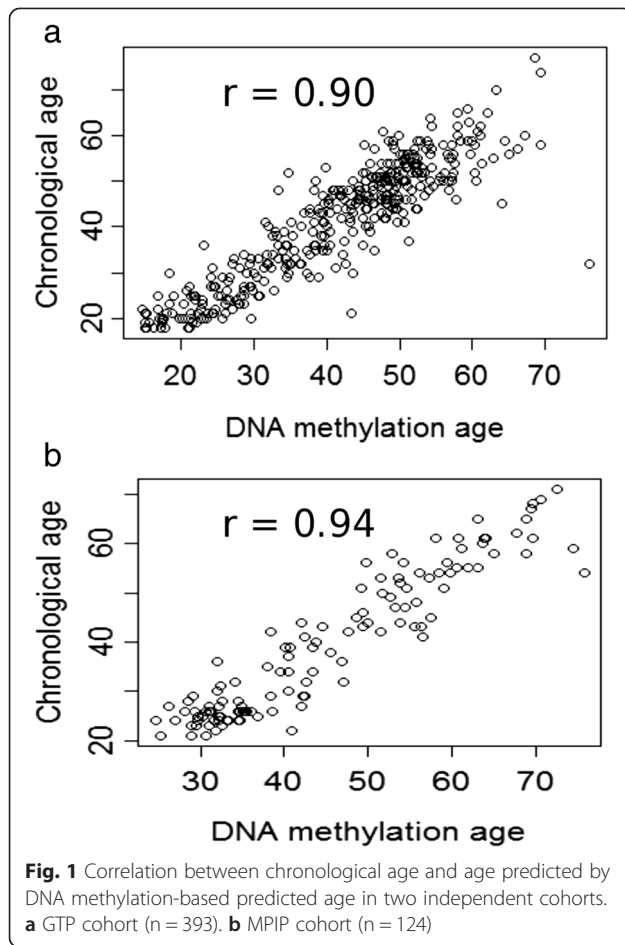
Results

Prediction of chronological age using the epigenetic clock

DNAM-age was calculated from peripheral blood from two independent samples, derived from the Grady Trauma Project (GTP) and the Max Planck Institute of Psychiatry (MPIP) cohorts using genome-wide Illumina HumanMethylation450 BeadChips (450 K), as previously described [27]. Given that the GTP primarily comprises (>90 %) African American participants, we excluded other ethnicities to minimize confounders. This resulted in a total of 393 participants with DNAM-age data. In contrast, the MPIP cohort consists only of Caucasian participants with a total of 124 participants with baseline DNAM-age data. The mean (SD, range) age was 41.33 years (12.85, range 18 to 77 years) for the GTP and 39.5 years (14.14, range 21 to 71 years) for the MPIP. The n (%) of female participants was 278 (70.7 %) for the GTP and 44 (35.5 %) for the MPIP. To validate the epigenetic clock predictor in our cohorts, we correlated DNAM-age with chronological age as previously described [27]. This correlation was strong for both the GTP ($r = 0.90$, $P < 2.2 \times 10^{-16}$) (Fig. 1a) and MPIP ($r = 0.94$, $P < 2.2 \times 10^{-16}$) cohorts (Fig. 1b) and proved robust and similar for both genders ($r = 0.89$ for male vs. $r = 0.90$ for female in the GTP; $r = 0.95$ for male vs. $r = 0.94$ for female in the MPIP).

Epigenetic age acceleration is associated with cumulative lifetime stress, but not childhood or current stress alone, in an urban, African American cohort

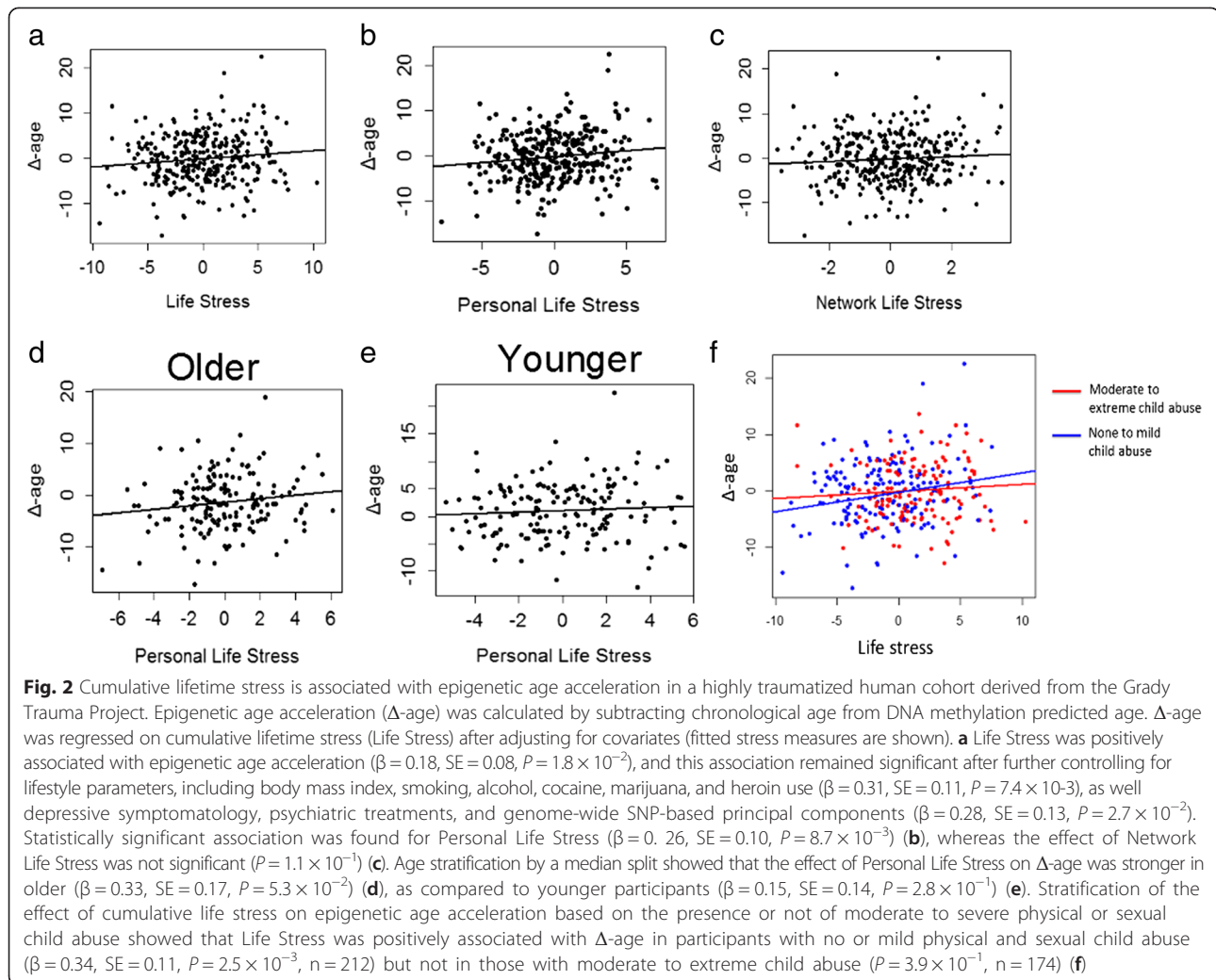
We then hypothesized that epigenetic age acceleration (Δ -age), calculated by subtracting the actual chronological



age from DNAM-age [27], would be positively associated with exposure to life stress. This hypothesis was tested in the highly traumatized GTP cohort. The mean (SD, range) Δ -age in the GTP was -0.13 years (5.69, range -17.31 to 43.98 years). A total of 304 GTP participants had data on lifetime stressors assessed by the Stressful Events Questionnaire (SEQ) and 386 participants had data on childhood maltreatment assessed by the Childhood Trauma Questionnaire (CTQ). The individual items from the SEQ were summed to yield a total score of lifetime stress exposure (Life Stress), and a similar total score was generated for the CTQ (Child Stress). The SEQ additionally assesses stressor exposure over the last year, and these items were summed to yield a score of more recent stress exposure (Current Stress). Linear regression models controlling for sex and age showed that Life Stress was positively associated with Δ -age ($\beta = 0.24$, $SE = 0.08$, $P = 2.8 \times 10^{-3}$), and this effect remained significant after further controlling for Houseman blood cell counts and technical batch effects ($\beta = 0.18$, $SE = 0.08$, $P = 1.8 \times 10^{-2}$) (Fig. 2a), lifestyle parameters, including body mass index, smoking,

alcohol, cocaine, marijuana, and heroin use ($\beta = 0.31$, $SE = 0.11$, $P = 7.4 \times 10^{-3}$), as well depressive symptomatology, psychiatric treatments, and genome-wide SNP-based principal components ($\beta = 0.28$, $SE = 0.13$, $P = 2.7 \times 10^{-2}$).

In secondary analyses, we examined whether the effect of lifetime stress on age acceleration depends on the type of stressor and other moderating variables. Based on previous work distinguishing between life events that affect the individual directly vs. life events that affect one's social network [42], we separately summed SEQ items assessing personal life events (Personal Life Stress) and items assessing network events (Network Life Stress). Δ -age showed a positive and significant association with Personal Life Stress ($\beta = 0.26$, $SE = 0.10$, $P = 8.7 \times 10^{-3}$) (Fig. 2b) and a positive but not significant association with Network Life Stress ($P = 1.1 \times 10^{-1}$) (Fig. 2c). No significant interactions were noted between Life Stress or Personal Life Stress and either sex or age. However, stratification of the GTP by a median split of age showed that the effect of Personal Life Stress on Δ -age was marginally stronger in older ($\beta = 0.33$, $SE = 0.17$, $P = 5.3 \times 10^{-2}$) (Fig. 2d) as compared to younger participants ($\beta = 0.15$, $SE = 0.14$, $P = 2.8 \times 10^{-1}$) (Fig. 2e). On the other hand, Δ -age was not associated with either CTQ score ($P = 4 \times 10^{-1}$) or Current Stress alone ($P = 1.3 \times 10^{-1}$). However, when participants were stratified based on the severity of childhood maltreatment, only individuals exposed to lower levels (none or mild) of sexual and physical childhood abuse (based on respective CTQ subscale scores) showed significant effects of Life Stress on Δ -age (Fig. 2f). This was not a consequence of differential stress exposure burden between the two groups, since, as expected, individuals exposed to higher levels of childhood abuse also had higher levels of Life Stress with a mean (SD) Life Stress of 12.32 (3.64) as compared to 10.01 (3.76) in individuals with lower levels of childhood abuse ($t_{299} = 5.38$, $P = 1.5 \times 10^{-7}$). Furthermore, the two strata showed similar correlations between DNAM-age and chronological age ($r = 0.91$ for higher vs. 0.92 for lower abuse, Fisher z score = 0.6, $P = 5.5 \times 10^{-1}$). Lastly, we found no association between Δ -age and current stress-related psychiatric phenotypes, including depressive ($P = 3.4 \times 10^{-1}$) and PTSD symptomatology ($P = 7.9 \times 10^{-1}$) in the GTP. In line with this finding, depression diagnosis was not associated with Δ -age in the MPIP cohort ($P = 2.3 \times 10^{-1}$, $n = 72$ controls vs. 52 depressed). Taken together, these findings show that cumulative lifetime stress, but not childhood trauma or current stress alone, is associated with accelerated epigenetic aging, an effect that is primarily driven by personal life events, may be more evident in advancing ages, and is blunted in participants exposed to high levels of childhood abuse.

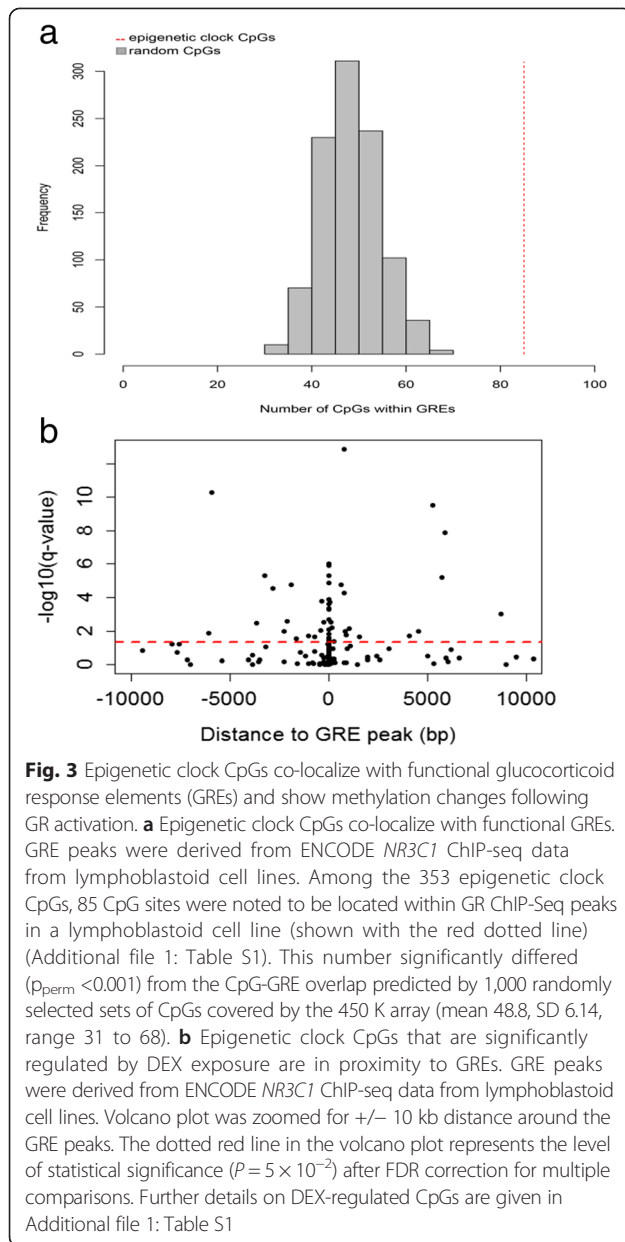


Epigenetic clock CpGs and neighboring genes are regulated by GR activation and show enriched association with aging-related diseases

The effect of lifetime stress on epigenetic aging prompted us to examine susceptibility of individual epigenetic clock CpGs to glucocorticoids, a primary molecular effector of stress responses, as a potential mechanism underlying this association. To address this hypothesis, we first examined whether epigenetic clock CpGs show DNA methylation changes 3 h after oral exposure to a GR agonist (1.5 mg of DEX) in the independent MPIP cohort ($n = 124$). After correcting for multiple testing, 110 of the 353 CpGs showed statistically significant methylation changes (false discovery rate (FDR)-adjusted $P < 5 \times 10^{-2}$). Among the DEX-regulated CpGs, 98 (89 %) showed decrease in methylation, whereas 12 (11 %) showed increase in methylation (Additional file 1: Table S1). We next examined the effect of acute DEX exposure on the epigenetic clock by comparing DNAM-age at baseline vs. 3 h after DEX exposure ($n = 124$). There was no effect

of DEX on DNA methylation-predicted age (baseline mean DNAM-age = 45.24 vs. post-DEX mean DNAM-age = 45.15, paired $t_{123} = 0.31$, $P = 7.6 \times 10^{-1}$).

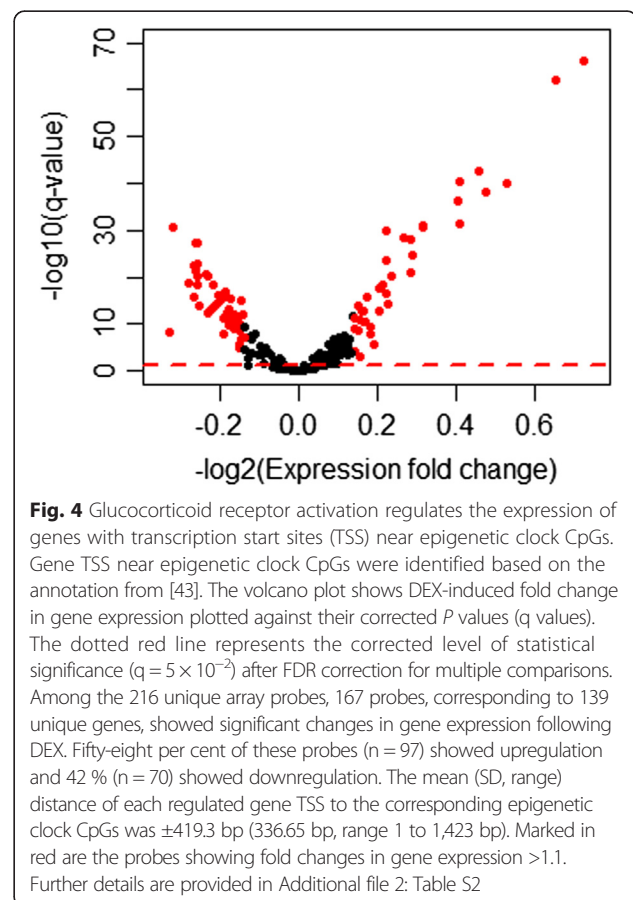
Given that GR binding to GREs can exert changes in DNA methylation, we then examined whether epigenetic clock CpGs co-localize with GREs. Among the 353 epigenetic clock CpGs, 85 CpGs were located within GREs as defined by CHIP-Seq peaks in a lymphoblastoid cell line (LCL) (Additional file 1: Table S1). This CpG-GRE co-localization significantly differed from the one expected by chance as determined by randomly drawing 1,000 sets ($n = 353$ CpGs) of CpG sites from all CpGs present on the 450 K array (expected mean 48.8, SD 6.1, range 31 to 68, $p_{\text{perm}} < 1 \times 10^{-3}$) (Fig. 3a). Proximity to GREs was particularly observed for DEX-regulated CpGs (Fig. 3b), with 17 of these sites located right within GREs and 35 within 1 kb distance from GREs. Because the 353 CpGs were originally derived from the 21,369 (21 K) CpGs that overlap the 27 K and 450 K Illumina arrays [27], we next examined whether the epigenetic clock



CpG-GRE co-localization differs from the one present in the 21 K background. Epigenetic clock CpG-GRE co-localization did not differ from the one expected by chance when randomly drawing 1,000 CpG sets ($n = 353$ CpGs) from the 21 K CpG sites (expected mean 3,094, SD 50.7, range 2,927–3,270, $p_{\text{perm}} = 9.7 \times 10^{-1}$). Given that this lack of enrichment could be the result of high CpG-GRE co-localization already present in the 21 K, as a last step we compared the co-localization present in the 21 K with the 450 K background and we noted significantly higher CpG-GRE co-localization in the 21 K as compared to the 450 K background ($p_{\text{perm}} < 1 \times 10^{-3}$). These findings suggest that the increased epigenetic clock CpG-GRE co-localization is a more general

property of the 21 K CpGs used to develop the epigenetic clock. Yet the presence of a high number of epigenetic clock CpGs within functional GREs is in line with our hypothesis that these sites may be highly susceptible to GR activation.

We then assessed whether genes that have transcription start sites (TSS) in the proximity of epigenetic clock CpGs are also dynamically regulated by GR activation. For this purpose, we used peripheral blood genome-wide gene expression array data in the MPIP cohort to examine the DEX-induced changes in the expression of genes with transcription start sites (TSS) close to epigenetic clock CpGs based on the 450 K annotation from [43]. Using these criteria, we annotated 344 unique genes. Of these, 333 genes were present on the gene expression microarray and a total of 170 genes, corresponding to 220 epigenetic clock CpGs, were expressed above background in the MPIP cohort (Additional file 2: Table S2). Transcription of these genes was detected by 216 unique gene expression array probes. After FDR-based correction for multiple testing, 167 out of the 216 detected probes, corresponding to 139 unique genes (81.7 %), showed significant changes in gene expression following DEX exposure (FDR-adjusted P values < 0.05) (Fig. 4).



Fifty-eight per cent of these probes ($n = 97$) showed up-regulation and 42 % ($n = 70$) showed downregulation. The mean (SD, range) distance of each regulated gene TSS to the corresponding epigenetic clock CpGs was ± 419.3 bp (336.65 bp, range 1 to 1,423 bp). To rule out potential bias derived from the 21 K background, we then asked whether genes neighboring epigenetic clock CpGs are more responsive to GR activation compared to genes neighboring the 21 K CpGs. A total of 5,443 unique genes, corresponding to 21,015 21 K CpGs, showed significant DEX-induced mRNA expression changes (FDR-adjusted P values $< 5 \times 10^{-2}$). The number of DEX-regulated genes was significantly higher for the genes with TSS close to epigenetic clock CpGs as compared to 21 K CpGs (Fisher's exact test $P = 6.3 \times 10^{-5}$). Taken together, these data demonstrate enhanced responsiveness of genes neighboring epigenetic clock CpGs to GR activation.

Lastly, we performed disease enrichment analysis in WebGestalt using the set of unique DEX-regulated genes ($n = 139$) as the input for the analysis and the genes expressed above background in our peripheral blood gene expression arrays as the reference set of genes. After FDR correction for multiple testing, this resulted in enriched association for aging-related diseases, including coronary artery disease, arteriosclerosis, and leukemias (FDR-adjusted $P < 5 \times 10^{-2}$ each) (Additional file 3: Table S3).

Discussion

The present study sought to determine the effect of life stressors on epigenetic aging, as measured with the epigenetic clock [27] in peripheral blood samples. While previous studies found associations of the epigenetic clock with several phenotypes [27, 35–41], this is the first study to use this predictor in a highly traumatized cohort. As hypothesized, accelerated epigenetic aging was associated with cumulative lifetime stress burden. Given that epigenetic effects of the stress response can be mediated by GR signaling, we further examined the molecular basis of this association by annotating epigenetic clock CpG sites in relation to GREs and examining the impact of GR activation on these sites. We found that GREs co-localize with epigenetic clock CpGs and that glucocorticoid activation can induce dynamic methylation changes of these sites as well as changes in the expression of genes neighboring epigenetic clock CpGs. Taken together, these converging findings support a model of stress-induced accelerated epigenetic aging, plausibly mediated by the lasting effects of cumulative stressor exposure and aberrant glucocorticoid signaling on the epigenome.

Further examination of the relationship between life stress and epigenetic aging led us to several interesting

observations. First, this relationship was apparent for cumulative stress exposure throughout the lifetime, whereas no significant association was found with childhood maltreatment or current stress alone. This finding is in accordance with a recent study observing no effect of childhood trauma on epigenetic aging in combat veterans [35] and suggests that cumulative stressors over the lifetime, rather than time-limited stressors either during childhood or adulthood, have a stronger or more lasting effect on epigenetic aging. Nonetheless, it is also possible that these null findings may be due to lack of power, the timing of DNA methylation assessments, or reversibility of epigenetic aging, possibilities that could be addressed by future longitudinal studies. Second, the effect of lifetime stress was driven by personal stressors – affecting the participant directly – rather than network stressors that occur to someone within the participant's network. This is congruent with previous studies showing that personal life events are more strongly correlated with genetic factors as compared to network events [44]. In line with the effects of lifetime vs. current stress, these effects were more pronounced in older individuals, suggesting cumulative epigenetic vulnerability in older individuals. Lastly, the epigenetic effects of lifetime stress were blunted in individuals with higher levels of childhood abuse. This finding could not be attributed to differences in the levels of lifetime stress, since individuals exposed to higher levels of childhood abuse also had higher levels of cumulative lifetime stress burden. Thus, it is possible that early trauma exposure triggers additional mechanisms of risk and resilience that may interfere with subsequent effects of stressors on epigenetic aging, a hypothesis that remains to be tested by future studies.

The effects of lifetime stress on epigenetic aging in peripheral blood are likely mediated by persistent neuroendocrine alterations induced by cumulative stress exposure. Stressors and glucocorticoids can drive persistent changes in the expression of glucocorticoid-responsive genes and concomitant changes in DNA methylation at CpGs located at or near GREs [17, 18, 22, 45]. Supporting this hypothesis, we noted that a high number of epigenetic clock CpG sites are located within functional GREs and show dynamic methylation changes following DEX exposure. Notably, most of these CpGs show DEX-induced decrease in methylation, whereas far fewer sites show increased methylation (98 vs. 12). This is in accordance with previous studies showing that activation of the GR results in local demethylation of CpGs in the proximity of a GRE [18, 22, 23] and that site-specific decreases in methylation have been implicated in aging-related phenotypes [46]. CpG demethylation has been proposed to be potentially mediated by at least two enzymatic processes, base excision repair and

oxidation [47, 48]. Examining the role of these processes may provide further insights into mechanisms of stress-induced epigenetic aging. Furthermore, an open question concerns the sequence of molecular events that determine whether some stress-induced DNA methylation changes become embedded and longlasting, while other changes are dynamic and reversible. Given the low dose and acute exposure to glucocorticoids in our study, additional experiments with different doses and more chronic *in vitro* or *in vivo* GR activation will be necessary to better elucidate this mechanism.

An important implication of our findings is the potential role of stress-induced epigenetic aging in health and disease. Increasing age and aging-related diseases have been associated with global and site-specific changes in DNA methylation [25–30, 39]. The age-related epigenetic clock CpGs co-localize with genes that show enrichment for cell growth and survival, organismal development, and cancer [27]. Furthermore, we show that DEX-regulated genes neighboring epigenetic clock CpGs show enriched association for aging-related diseases, including coronary artery disease, arteriosclerosis, and leukemias. These findings raise the possibility that lifetime stress may contribute to these diseases via its cumulative impact on epigenetic regulation of genes implicated in aging-related diseases.

The findings of the present study should be viewed in the context of its limitations. Although we observe an association between epigenetic age acceleration and lifetime stressors in the GTP cohort, the cross-sectional design of the study limits conclusions regarding the direction of causality. As discussed above, it is plausible that epigenetic aging of peripheral blood cells results from persistent alterations of the neuroendocrine but also immune milieu induced by repetitive stressor exposure. However, accelerated epigenetic aging might alternatively represent a vulnerability marker that predisposes individuals to expose themselves to stressful environments. It is also important to acknowledge that, while the high levels of traumatic events in the GTP make this cohort highly suitable for examining the effects of lifetime stress on the epigenome, they may also limit generalizability of these findings to other less traumatized cohorts. Moreover, the present study examined epigenetic aging in peripheral blood only. While this tissue is easily accessible and relevant for biomarker research, other tissues may be more susceptible to psychological stress and should be examined in the context of specific diseases. For example, disease-specific effects on the epigenetic clock have been demonstrated for liver tissue in the context of obesity [39]. Another limitation is the use of Chip-Seq data from lymphoblastoid cell lines to examine epigenetic clock CpG-GRE co-localization. This cell line represents the best available proxy for peripheral blood, the source

tissue for our methylation data, but this approach may also be limited by the tissue specificity of functional GREs and the altered epigenetic landscapes of immortalized cell lines. Lastly, although we corrected for several confounders that might influence DNA methylation, such as sex, age, smoking, body mass index, substance abuse, current psychiatric symptoms and treatments, other factors not captured by our methods may have confounded the observed relationships. These limitations may be overcome in future studies by employing detailed prospective measurements of lifestyle factors, stressor exposure, DNA methylation, and incidence of stress-related phenotypes at different time points throughout the lifetime.

Conclusions

The present study provides evidence that cumulative life stress exposure is associated with accelerated epigenetic aging and that these effects may be mediated by glucocorticoid signaling. Our findings further suggest that DNA methylation-based age prediction in peripheral blood may be a useful molecular marker to incorporate in future studies examining the effects of life stress exposure. These findings offer novel insights into the molecular mechanisms linking psychological stress with diseases of the aging.

Methods

Clinical samples

The effect of lifetime stress on epigenetic aging was examined in the Grady Trauma Project (GTP), a large study conducted in Atlanta, Georgia, that investigates the role of genetic and environmental factors in shaping responses to stressful life events. The GTP includes more than 7,000 participants from a predominantly African American, urban population of low socioeconomic status [49, 50]. This population is characterized by high prevalence and severity of trauma over the lifetime and is thus particularly relevant for examining the effects of stressors on epigenetic markers. For this purpose, we used a subsample of GTP participants with genome-wide DNA methylation data. All participants provided written informed consent and all procedures were approved by the Institutional Review Boards of the Emory University School of Medicine and Grady Memorial Hospital (IRB00002114).

We examined glucocorticoid-induced methylation changes of epigenetic clock CpGs and responsivity of genes closest to these CpGs in 297 Caucasian participants recruited at the Max Planck Institute of Psychiatry (MPIP). Recruitment strategies and characterization of participants have been previously described [51, 52]. These consisted of 200 male (83 healthy probands and 117 inpatients with depressive disorders) and 97 female (48 healthy probands and 49 depressed) individuals. Baseline whole blood samples were obtained at 18:00 after 2 h

of fasting and abstention from coffee and physical activity (baseline). Participants then received 1.5 mg oral dexamethasone (DEX) and a second blood draw was performed at 21:00, 3 h after DEX ingestion (post-DEX). The study was approved by the local ethics committee (approval number: 318/00) and all individuals gave written informed consent. All experimental methods comply with the Helsinki Declaration.

Psychometric instruments

Childhood trauma was measured in the GTP with the Childhood Trauma Questionnaire (CTQ), a validated self-report questionnaire that assesses five types of maltreatment during childhood: sexual, physical, and emotional abuse, as well as emotional and physical neglect [53]. Scores for each type of maltreatment were derived from participant responses to questionnaire items and scores from all types were summed to yield a total CTQ score reflecting overall burden of childhood maltreatment. Moderate to extreme sexual abuse was defined by a cutoff score of 8 or above in the CTQ sexual abuse subscale, and moderate to extreme physical abuse was defined by a cutoff score of 10 or above in the physical abuse subscale as previously described [54].

Stressful lifetime events in the GTP were assessed with the Stressful Events Questionnaire (SEQ), a 39-item self-report instrument that has been described in detail [55]. The SEQ covers a wide range of stressor exposure, ranging from personal life events, such as divorce, unemployment, crime, and financial stressors, to network life events, such as knowing someone who was murdered. Participants report whether they have experienced these events either in the past year or at any time in their life. Although the SEQ assesses life event exposure throughout the lifetime, it does not include questions specific for childhood maltreatment. Life events are summed to yield a total score that reflects the number of stressors experienced over the last year (Current Stress) or cumulative number of stressors experienced throughout one's lifetime (Life Stress).

Participants underwent the Structured Clinical Interviews for DSM-IV defined psychiatric diagnoses. Given the observed relation between stress-related psychiatric disorders and accelerated cellular aging, we also examined major depression and PTSD as variables of interest. In the GTP, current depressive symptomatology was assessed with the 21-item validated Beck Depression Inventory (BDI) [56, 57] and current PTSD symptomatology was assessed with the validated 17-item PTSD Symptom Scale (PSS) [49, 58].

DNA methylation

Genomic DNA from the GTP cohort ($n = 393$) and the MPIP ($n = 124$) was extracted from whole blood using

the Gentra Puregene Blood Kit (QIAGEN). DNA quality and quantity was assessed by NanoDrop 2000 Spectrophotometer (Thermo Scientific) and Quant-iT Picogreen (Invitrogen). Genomic DNA was bisulfite converted using the Zymo EZ-96 DNA Methylation Kit (Zymo Research) and DNA methylation levels were assessed for >480,000 CpG sites using the Illumina Human-Methylation450 BeadChip array. Hybridization and processing was performed according to manufacturer's instructions as previously described [59]. Quality control of methylation data, including intensity read outs, filtering (detection P value >0.01 in at least 75 % of the samples), cellular composition estimation, as well as beta and M-value calculation was done using the minfi Bioconductor R package version 1.10.2 [60].

For the GTP cohort, X chromosome, Y chromosome, and non-specific binding probes were removed [61]. We also excluded probes if single nucleotide polymorphisms (SNPs) were documented in the interval for which the Illumina probe is designed to hybridize. Given that the GTP cohort includes individuals from different ethnicities, we also removed probes if they were located close (10 bp from query site) to a SNP which had Minor Allele Frequency of ≥ 0.05 , as reported in the 1,000 Genomes Project, for any of the populations represented in the samples. Technical batch effects were identified by inspecting the association of the first principal components of the methylation levels with plate, sentrix array, and position (row) and by further visual inspection of principal component plots using the shinyMethyl Bioconductor R package version 0.99.3 [62]. This procedure identified row and slide as technical batches. The raw methylation data and all related phenotypes for the GTP cohort have been deposited into NCBI GEO (GSE72680).

For the MPIP cohort, filtered beta values were reduced by eliminating any CpG sites/probes on sex chromosomes, as well as probes found to have SNPs at the CpG site itself or in the single-base extension site with a MAF ≥ 1 % in the 1,000 Genomes Project European population and/or non-specific binding probes according to [61]. Additionally, we performed a re-alignment of the array probe sequences using Bismark (doi: 10.1093/bioinformatics/btr167). This yielded a total of 425,883 CpG sites for further analysis. Using the same procedure for batch identification as above, we identified processing (experiment) date as technical batch in the MPIP. The data were then normalized with functional normalization [63], an extension of quantile normalization included in the minfi R package and batch-corrected using ComBat. The raw methylation data and all related phenotypes for the MPIP cohort have been deposited into NCBI GEO (GSE74414).

Gene expression

In the DEX-treated (MPIP) cohort ($n = 297$, including the 124 individuals used for the MPIP methylation analysis), both baseline and post-DEX whole blood RNA was collected using PAXgene Blood RNA Tubes (PreAnalytiX), processed as described previously [51, 52]. Samples had a mean RNA integrity number (RIN) of 8 ± 0.51 SD. Blood RNA was hybridized to Illumina HumanHT-12 v3 and v4 Expression BeadChips (Illumina, San Diego, CA, USA). Raw probe intensities were exported using Illumina's GenomeStudio and further statistical processing was carried out using R. All 29,075 probes present on both microarrays, excluding X and Y chromosomes as well as cross-hybridizing probes identified by using the Re-Annotator pipeline (<http://dx.doi.org/10.1101/019596>) were first filtered with an Illumina detection P value of 0.05 in at least 50 % of the samples, leaving 11,994 expressed probes for further analysis. Subsequently, each transcript was transformed and normalized through variance stabilization and normalization (VSN) [64]. Using the same procedure for batch identification as for the methylation data, we identified slide, amplification round, array version, and amplification plate column as technical batches. The data were then adjusted using ComBat [65] and have been deposited into NCBI GEO (GSE64930).

Statistical analyses

All statistical analyses were conducted in R version 3.1.0 (<http://www.r-project.org/>) [66]. Unless indicated otherwise, P values are nominal and two-tailed. All corrections for multiple testing were performed using the FDR method of Benjamini and Hochberg. The level of statistical significance was set a priori at 0.05 (5×10^{-2}).

DNA methylation-based age prediction was performed using the R code and statistical pipeline developed by Horvath [27]. This predictor was developed using 82 Illumina DNA methylation array datasets ($n = 7,844$) involving 51 healthy tissues and cell types [27]. The raw data were normalized using BMIQ normalization method [67] implemented in the Horvath DNA methylation-based age predictor R script [27]. Robustness and reproducibility of the epigenetic age predictor was tested using 40× technical replicates of an individual control sample, randomized across microarray chips and batches used to measure DNA methylation in the GTP cohort. The average epigenetic age (DNAM-age) of the control sample (true age = 32 years) was 32.64 (SD: 0.23) years with an average correlation $r = 0.97$ (0.001). Age acceleration (Δ -age) was defined (as previously) as the average difference between DNAM-age and chronological age. One GTP participant had extreme Δ -age (43.98 years), and using the Grubbs' test (<http://graphpad.com/quickcalcs/grubbs2/>) was noted to be the

only outlier ($Z = 3.80$, $P < 5 \times 10^{-2}$). Although primary analyses were conducted without this outlier, inclusion of this individual did not substantially alter the reported results. Generalized linear regression models tested the relationship of Δ -age with stressors and stress-related phenotypes (GTP cohort). Because DNAM-age is calculated from raw beta values (before Combat correction for batches), technical batches identified for the GTP (row and slide) and the MPIP cohort (processing date) were tested as potential confounders in the respective regression models. In the GTP, models were further adjusted for age, sex, Houseman cell counts, body mass index, smoking, alcohol, current substance abuse, and the principal components from population stratification checks. In the MPIP, models were adjusted for gender, age, body mass index, and Houseman cell counts.

To determine if methylation signals or gene expression levels are significantly different before and after DEX stimulation in the MPIP cohort, likelihood ratio tests accounting for gender, age, body mass index, disease status, and estimated cell-type counts were applied to each CpG site ($n = 353$) and expression array probe ($n = 11,994$), respectively. DNA methylation and gene expression changes were corrected for multiple comparisons using FDR. The 353 epigenetic clock CpGs were annotated to a total of 344 genes. Among these, 170 genes were detected in peripheral blood by 216 gene expression array probes (163 genes were expressed below background and 11 genes were not covered by the gene expression arrays).

To account for population stratification due to discrepancies between self-reported and actual race in the GTP, we used genome-wide SNP data that were available for 382 participants. Of the 700 k SNPs present on the Omni Quad and Omni express arrays, 645,8315 autosomal SNPs were left after filtering with the following criteria: minor allele frequency of $>1\%$; Hardy-Weinberg equilibrium of 0.000001; and genotyping rate of $>98\%$. The samples were clustered to calculate rates of identity by descent (IBD). We then ran multidimensional scaling analysis on the IBD matrix using PLINK2 (<https://www.cog-genomics.org/plink2>) and plotted the first ten axes of variation against each other. No outliers were detected. The first two principal components were used as covariates in regression models to adjust for population stratification.

To identify whether epigenetic clock CpG sites are co-localized with GREs, we used ENCODE NR3C1 ChIP-Seq data from lymphoblastoid cell lines (*accession: ENCSR904YPP*) for which no aligned tracks are currently available. Initial filtering was performed using FASTX Toolkit (v. 0.0.14, http://hannonlab.cshl.edu/fastx_toolkit/index.html) and Prinseq (v. 0.20.3) [68] to eliminate artefacts and low quality reads. Alignment on

hg19 was performed using BWA (v. 0.7.10) [69] allowing only uniquely mappable alignments with an alignment quality of above 20. Reads from both ChIP-Seq and both control libraries were pooled leading to 46,453,650 and 68,227,580 used reads, respectively. Peak-calling was carried out by MACS14 (v. 1.4.2) [70] using default settings, resulting in approximately 23,000 annotated signals. The average length of ChIP-Seq signal as defined by the peak calling was 746.3 bps (SD: 370.6). We generated 1,000 sets ($n = 353$ CpGs) of randomly drawn CpG sites (without replacement) from the set of all CpGs present on the 450 K BeadChip array (excluding X and Y chromosomes). For every set we counted the percentage of CpG sites within a GRE ChIP-Seq signal (± 0 bp). On this basis we constructed the null distribution and compared it to the observed percentage of clock CpG sites within a GRE ChIP-Seq signal to measure the enrichment statistics.

Disease enrichment analysis was performed using the WEB-based GENE SeT ANALYSIS Toolkit (WebGestalt; <http://bioinfo.vanderbilt.edu/webgestalt/>) [71, 72]. This was performed by using as input the set of unique DEX-regulated genes neighboring epigenetic clock CpGs ($n = 139$) and as reference the set of genes expressed above background in our peripheral blood gene expression arrays. The minimum number of genes for the enrichment analysis was set at 5, the statistic performed was hypergeometric test, and results were corrected for multiple testing using FDR.

Additional files

Additional file 1: Table S1. Location of epigenetic clock CpGs in relation to the nearest glucocorticoid response element (as shown by within GR ChIP-Seq peaks in a lymphoblastoid cell line) and their methylation changes in response to the glucocorticoid receptor agonist dexamethasone. (DEX). (XLSX 68 kb)

Additional file 2: Table S2. Annotation of genes with transcription start sites (TSS) near epigenetic clock CpGs and their expression changes in response to DEX. Gene annotation was based on [43]. (XLSX 26 kb)

Additional file 3: Table S3. WebGestalt Disease enrichment analysis of the set of unique DEX-regulated genes ($n = 139$) with TSS near epigenetic clock CpGs. For the primary analysis, we used as reference the set of genes expressed above background in our peripheral blood gene expression arrays. This analysis was repeated using a more condensed background comprised only of the genes neighboring 21 K CpGs that showed DEX-induced mRNA expression changes ($n = 5,443$). While this post-hoc analysis yielded no statistically significant results after correction for multiple testing (P values presented in the last column), the top 10 diseases were very similar (with higher but nominally significant P values for the top three hits) with the analysis using the broader reference set of genes. (XLSX 10 kb)

Competing interests

The authors declare no competing interests.

Authors' contributions

ASZ, EBB, and DM conceived and designed the study. TCR, JA, and SI were involved in the DNA methylation analyses. JA, SR, and AM analyzed the DEX-treated sample. JA and SR performed the GRE enrichment analysis. KJR, AKS, BB, JFL, MI, TB, AE, CBH, and CH contributed the clinical samples. ASZ

performed the statistical analyses with substantial input from JA, TCR, NRW, and DM. ASZ performed the disease enrichment analysis with input from JA, EBB, and DM. ASZ wrote the manuscript with input from EBB and DM. All authors read and approved the final manuscript.

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References

- Centers for Disease Control and Prevention. The State of Aging and Health in America 2013. Atlanta, GA: Centers for Disease Control and Prevention; 2013.
- Office for National Statistics. National Population Projections, 2012-based Statistical Bulletin. Newport: Office for National Statistics; 2012.
- Niccoli T, Partridge L. Ageing as a risk factor for disease. *Curr Biol*. 2012;22:R741–52.
- Karatsoreos IN, McEwen BS. Psychobiological allostasis: resistance, resilience and vulnerability. *Trends Cogn Sci*. 2011;15:576–84.

5. Epel ES, Blackburn EH, Lin J, Dhabhar FS, Adler NE, Morrow JD, et al. Accelerated telomere shortening in response to life stress. *Proc Natl Acad Sci U S A*. 2004;101:17312–5.
6. Epel ES, Lin J, Wilhelm FH, Wolkowitz OM, Cawthon R, Adler NE, et al. Cell aging in relation to stress arousal and cardiovascular disease risk factors. *Psychoneuroendocrinology*. 2006;31:277–87.
7. Chandola T, Brunner E, Marmot M. Chronic stress at work and the metabolic syndrome: prospective study. *BMJ*. 2006;332:521–5.
8. Rozanski A, Blumenthal JA, Kaplan J. Impact of psychological factors on the pathogenesis of cardiovascular disease and implications for therapy. *Circulation*. 1999;99:2192–217.
9. Powell ND, Allen RG, Hufnagle AR, Sheridan JF, Bailey MT. Stressor-induced alterations of adaptive immunity to vaccination and viral pathogens. *Immunol Allergy Clin North Am*. 2011;31:69–79.
10. Powell ND, Tarr AJ, Sheridan JF. Psychosocial stress and inflammation in cancer. *Brain Behav Immun*. 2013;30(Suppl):S41–7.
11. Zannas AS, McQuoid DR, Steffens DC, Chrousos GP, Taylor WD. Stressful life events, perceived stress, and 12-month course of geriatric depression: direct effects and moderation by the 5-HTTLPR and COMT Val158Met polymorphisms. *Stress*. 2012;15:425–34.
12. Peavy GM, Salmon DP, Jacobson MW, Hervey A, Gamst AC, Wolfson T, et al. Effects of chronic stress on memory decline in cognitively normal and mildly impaired older adults. *Am J Psychiatry*. 2009;166:1384–91.
13. Danese A, McEwen BS. Adverse childhood experiences, allostasis, allostatic load, and age-related disease. *Physiol Behav*. 2012;106:29–39.
14. Felitti VJ, Anda RF, Nordenberg D, Williamson DF, Spitz AM, Edwards V, et al. Relationship of childhood abuse and household dysfunction to many of the leading causes of death in adults. The Adverse Childhood Experiences (ACE) Study. *Am J Prev Med*. 1998;14:245–58.
15. Danese A, Moffitt TE, Pariante CM, Ambler A, Poulton R, Caspi A. Elevated inflammation levels in depressed adults with a history of childhood maltreatment. *Arch Gen Psychiatry*. 2008;65:409–15.
16. Vaccarino V, Goldberg J, Rooks C, Shah AJ, Veledar E, Faber TL, et al. Post-traumatic stress disorder and incidence of coronary heart disease: a twin study. *J Am Coll Cardiol*. 2013;62:970–8.
17. Zannas AS, West AE. Epigenetics and the regulation of stress vulnerability and resilience. *Neuroscience*. 2014;264:157–70.
18. Klengel T, Mehta D, Anacker C, Rex-Haffner M, Pruessner JC, Pariante CM, et al. Allele-specific FKBP5 DNA demethylation mediates gene-childhood trauma interactions. *Nat Neurosci*. 2013;16:33–41.
19. Weaver IC, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, et al. Epigenetic programming by maternal behavior. *Nat Neurosci*. 2004;7:847–54.
20. Chrousos GP, Gold PW. The concepts of stress and stress system disorders. Overview of physical and behavioral homeostasis. *JAMA*. 1992;267:1244–52.
21. Bamberger CM, Schulte HM, Chrousos GP. Molecular determinants of glucocorticoid receptor function and tissue sensitivity to glucocorticoids. *Endocr Rev*. 1996;17:245–61.
22. Thomasson H, Flavin M, Espinas ML, Grange T. Glucocorticoid-induced DNA demethylation and gene memory during development. *EMBO J*. 2001;20:1974–83.
23. Wiench M, John S, Baek S, Johnson TA, Sung MH, Escobar T, et al. DNA methylation status predicts cell type-specific enhancer activity. *EMBO J*. 2011;30:3028–39.
24. Wiench M, Miranda TB, Hager GL. Control of nuclear receptor function by local chromatin structure. *FEBS J*. 2011;278:2211–30.
25. Heyn H, Li N, Ferreira HJ, Moran S, Pisano DG, Gomez A, et al. Distinct DNA methylomes of newborns and centenarians. *Proc Natl Acad Sci U S A*. 2012;109:10522–7.
26. Bjornsson HT, Sigurdsson MI, Fallin MD, Irizarry RA, Aspelund T, Cui H, et al. Intra-individual change over time in DNA methylation with familial clustering. *JAMA*. 2008;299:2877–83.
27. Horvath S. DNA methylation age of human tissues and cell types. *Genome Biol*. 2013;14:R115.
28. Christensen BC, Houseman EA, Marsit CJ, Zheng S, Wrensch MR, Wiemels JL, et al. Aging and environmental exposures alter tissue-specific DNA methylation dependent upon CpG island context. *PLoS Genet*. 2009;5:e1000602.
29. Hernandez DG, Nalls MA, Gibbs JR, Arepalli S, van der Brug M, Chong S, et al. Distinct DNA methylation changes highly correlated with chronological age in the human brain. *Hum Mol Genet*. 2011;20:1164–72.
30. Rakyán VK, Down TA, Maslau S, Andrew T, Yang TP, Beyan H, et al. Human aging-associated DNA hypermethylation occurs preferentially at bivalent chromatin domains. *Genome Res*. 2010;20:434–9.
31. Horvath S, Zhang Y, Langfelder P, Kahn RS, Boks MP, van Eijk K, et al. Aging effects on DNA methylation modules in human brain and blood tissue. *Genome Biol*. 2012;13:R97.
32. Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sada S, et al. Genome-wide methylation profiles reveal quantitative views of human aging rates. *Mol Cell*. 2013;49:359–67.
33. Weidner CI, Lin Q, Koch CM, Eisele L, Beier F, Ziegler P, et al. Aging of blood can be tracked by DNA methylation changes at just three CpG sites. *Genome Biol*. 2014;15:R24.
34. Bocklandt S, Lin W, Sehl ME, Sanchez FJ, Sinsheimer JS, Horvath S, et al. Epigenetic predictor of age. *PLoS One*. 2011;6:e14821.
35. Boks MP, Mierlo HC, Rutten BP, Radstake TR, De Witte L, Geuze E, et al. Longitudinal changes of telomere length and epigenetic age related to traumatic stress and post-traumatic stress disorder. *Psychoneuroendocrinology*. 2015;51:506–12.
36. Marioni RE, Shah S, McRae AF, Chen BH, Colicino E, Harris SE, et al. DNA methylation age of blood predicts all-cause mortality in later life. *Genome Biol*. 2015;16:25.
37. Marioni RE, Shah S, McRae AF, Ritchie SJ, Muniz-Terrera G, Harris SE, et al. The epigenetic clock is correlated with physical and cognitive fitness in the Lothian Birth Cohort 1936. *Int J Epidemiol*. 2015;44:1388–96.
38. Horvath S, Garagnani P, Bacalini MG, Pirazzini C, Salvioli S, Gentilini D, et al. Accelerated epigenetic aging in Down syndrome. *Aging Cell*. 2015;14:491–5.
39. Horvath S, Erhart W, Brosch M, Ammerpohl O, von Schonfels W, Ahrens M, et al. Obesity accelerates epigenetic aging of human liver. *Proc Natl Acad Sci U S A*. 2014;111:15538–43.
40. Miller GE, Yu T, Chen E, Brody GH. Self-control forecasts better psychosocial outcomes but faster epigenetic aging in low-SES youth. *Proc Natl Acad Sci U S A*. 2015;112:10325–30.
41. Kananen L, Nevalainen T, Jylhava J, Marttila S, Hervonen A, Jylha M, et al. Cytomegalovirus infection accelerates epigenetic aging. *Exp Gerontol*. 2015;72:227–9.
42. Kendler KS, Karkowski LM, Prescott CA. Causal relationship between stressful life events and the onset of major depression. *Am J Psychiatry*. 1999;156:837–41.
43. Price ME, Cotton AM, Lam LL, Farre P, Emberly E, Brown CJ, et al. Additional annotation enhances potential for biologically-relevant analysis of the Illumina Infinium HumanMethylation450 BeadChip array. *Epigenetics Chromatin*. 2013;6:4.
44. Bolinsky PK, Neale MC, Jacobson KC, Prescott CA, Kendler KS. Sources of individual differences in stressful life event exposure in male and female twins. *Twin Res*. 2004;7:33–8.
45. Lee RS, Tamashiro KL, Yang X, Purcell RH, Harvey A, Willour VL, et al. Chronic corticosterone exposure increases expression and decreases deoxyribonucleic acid methylation of Fkbp5 in mice. *Endocrinology*. 2010;151:4332–43.
46. Vandiver AR, Irizarry RA, Hansen KD, Garza LA, Runarsson A, Li X, et al. Age and sun exposure-related widespread genomic blocks of hypomethylation in nonmalignant skin. *Genome Biol*. 2015;16:80.
47. Niehrs C, Schafer A. Active DNA demethylation by Gadd45 and DNA repair. *Trends Cell Biol*. 2012;22:220–7.
48. Pastor WA, Aravind L, Rao A. TETonic shift: biological roles of TET proteins in DNA demethylation and transcription. *Nat Rev Mol Cell Biol*. 2013;14:341–56.
49. Binder EB, Bradley RG, Liu W, Epstein MP, Deveau TC, Mercer KB, et al. Association of FKBP5 polymorphisms and childhood abuse with risk of posttraumatic stress disorder symptoms in adults. *JAMA*. 2008;299:1291–305.
50. Gillespie CF, Bradley B, Mercer K, Smith AK, Conneely K, Gapen M, et al. Trauma exposure and stress-related disorders in inner city primary care patients. *Gen Hosp Psychiatry*. 2009;31:505–14.
51. Menke A, Arloth J, Putz B, Weber P, Klengel T, Mehta D, et al. Dexamethasone stimulated gene expression in peripheral blood is a sensitive marker for glucocorticoid receptor resistance in depressed patients. *Neuropsychopharmacology*. 2012;37:1455–64.
52. Arloth J, Bogdan R, Weber P, Frishman G, Menke A, Wagner KV, et al. Genetic differences in the immediate transcriptome response to stress predict risk-related brain function and psychiatric disorders. *Neuron*. 2015;86:1189–202.

53. Bernstein DP, Stein JA, Newcomb MD, Walker E, Pogge D, Ahluvalia T, et al. Development and validation of a brief screening version of the Childhood Trauma Questionnaire. *Child Abuse Negl.* 2003;27:169–90.
54. DiLillo D, Fortier MA, Hayes SA, Trask E, Perry AR, Messman-Moore T, et al. Retrospective assessment of childhood sexual and physical abuse: a comparison of scaled and behaviorally specific approaches. *Assessment.* 2006;13:297–312.
55. Smith AK, Conneely KN, Kilaru V, Mercer KB, Weiss TE, Bradley B, et al. Differential immune system DNA methylation and cytokine regulation in post-traumatic stress disorder. *Am J Med Genet B Neuropsychiatr Genet.* 2011;156B:700–8.
56. Beck AT, Steer RA, Garbin MG. Psychometric Properties of the Beck Depression Inventory - 25 Years of Evaluation. *Clinical Psychology Review.* 1988;8:77–100.
57. Beck AT, Ward CH, Mendelson M, Mock J, Erbaugh J. An inventory for measuring depression. *Arch Gen Psychiatry.* 1961;4:561–71.
58. Coffey SF, Dansky BS, Falsetti SA, Saladin ME, Brady KT. Screening for PTSD in a substance abuse sample: psychometric properties of a modified version of the PTSD Symptom Scale Self-Report. *Posttraumatic stress disorder. J Trauma Stress.* 1998;11:393–9.
59. Mehta D, Klengel T, Conneely KN, Smith AK, Altmann A, Pace TW, et al. Childhood maltreatment is associated with distinct genomic and epigenetic profiles in posttraumatic stress disorder. *Proc Natl Acad Sci U S A.* 2013;110:8302–7.
60. Aryee MJ, Jaffe AE, Corrada-Bravo H, Ladd-Acosta C, Feinberg AP, Hansen KD, et al. Minfi: a flexible and comprehensive Bioconductor package for the analysis of Infinium DNA methylation microarrays. *Bioinformatics.* 2014;30:1363–9.
61. Chen YA, Lemire M, Choufani S, Butcher DT, Grafodatskaya D, Zanke BW, et al. Discovery of cross-reactive probes and polymorphic CpGs in the Illumina Infinium HumanMethylation450 microarray. *Epigenetics.* 2013;8:203–9.
62. Fortin JP, Fertig E, Hansen K. shinyMethyl: interactive quality control of Illumina 450k DNA methylation arrays in R. *F1000Res.* 2014;3:175.
63. Fortin JP, Labbe A, Lemire M, Zanke BW, Hudson TJ, Fertig EJ, et al. Functional normalization of 450 k methylation array data improves replication in large cancer studies. *Genome Biol.* 2014;15:503.
64. Lin SM, Du P, Huber W, Kibbe WA. Model-based variance-stabilizing transformation for Illumina microarray data. *Nucleic Acids Res.* 2008;36:e11.
65. Johnson WE, Li C, Rabinovic A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics.* 2007;8:118–27.
66. R Development Core Team. *A Language and Environment for Statistical Computing.* Vienna: R Foundation for Statistical Computing; 2014.
67. Teschendorff AE, Marabita F, Lechner M, Bartlett T, Tegner J, Gomez-Cabrero D, et al. A beta-mixture quantile normalization method for correcting probe design bias in Illumina Infinium 450 k DNA methylation data. *Bioinformatics.* 2013;29:189–96.
68. Schmieder R, Edwards R. Quality control and preprocessing of metagenomic datasets. *Bioinformatics.* 2011;27:863–4.
69. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics.* 2009;25:1754–60.
70. Zhang Y, Liu T, Meyer CA, Eeckhoute J, Johnson DS, Bernstein BE, et al. Model-based analysis of ChIP-Seq (MACS). *Genome Biol.* 2008;9:R137.
71. Wang J, Duncan D, Shi Z, Zhang B. WEB-based GENE SeT Analysis Toolkit (WebGestalt): update 2013. *Nucleic Acids Res.* 2013;41:W77–83.
72. Zhang B, Kirov S, Snoddy J. WebGestalt: an integrated system for exploring gene sets in various biological contexts. *Nucleic Acids Res.* 2005;33:W741–8.

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